



ISSN 2414-438X (Print)
ISSN 2414-441X (Online)

THEORY AND PRACTICE OF MEAT PROCESSING

ТЕОРИЯ И ПРАКТИКА ПЕРЕРАБОТКИ МЯСА

2020, vol.5, no.3

ЦЕЛИ И ЗАДАЧИ ЖУРНАЛА

Приоритетной целью Журнала «Теория и практика переработки мяса» является распространение в мировом научном сообществе трудов по науке о мясе ученых научных центров, научно-исследовательских институтов и высших учебных заведений из России и стран СНГ, повышение уровня присутствия достижений представляемой ими науки на международной арене, знакомство Российских ученых с исследованиями за рубежом, освещение результатов перспективных направлений научно-исследовательской деятельности в мясной и птицеперерабатывающей промышленности.

К публикации в журнале приглашаются как отечественные, так и зарубежные ученые и специалисты.

Важнейшими задачами журнала являются: обобщение научных и практических достижений в области науки о мясе, повышения научной и практической квалификации как научных работников, так и представителей промышленности.

FOCUS AND SCOPE

The top priority goal of the Journal “Teoriâ i praktika pererabotki mâsa” (Theory and practice of meat processing) is to distribute in the world scientific community the results of the research in the field of meat science performed by the scientists from scientific centers, scientific-research institutes and institutions of higher education from Russia and the CIS countries, increase the level of presentation of the achievements of the respective science in the international arena, inform the Russian scientists about the research carried out abroad, highlight the results of the prospect directions of the research activities in the meat and poultry processing industries.

Both Russian and foreign scientists and experts are invited for publication in the journal.

The main tasks are generalization of scientific and practical achievements in the fields of meat science, increase scientific and practical qualifications as researchers and industry representatives.

РАЗДЕЛЫ ЖУРНАЛА

- ◆ Исследования в области биотехнологии, биохимии, физиологии, ланималогии, нутрициологии
- ◆ Исследование и создание новых технологий мясных производств
- ◆ Процессы, оборудование и аппараты мясных производств
- ◆ Стандартизация, сертификация, системы управления качеством и безопасность
- ◆ Микробиология, санитария и гигиена мяса и мясных продуктов
- ◆ Экономика
- ◆ Автоматизация и информатизация технологических процессов

SECTION POLICIES

- ◆ Investigations in the field of biotechnology, biochemistry, physiology, lanimology, nutritiology
- ◆ Investigation and development of new technologies of meat production
- ◆ Processes, equipment and apparatus of meat production
- ◆ Standardization, certification, systems of quality and safety management
- ◆ Microbiology, sanitary and hygiene of meat and meat products
- ◆ Economics
- ◆ Automation and informatization of the technological processes in the meat sect

**Министерство науки и высшего образования
Российской Федерации**
Minister of Science and Higher Education
of the Russian Federation

Федеральное государственное бюджетное
научное учреждение «Федеральный научный центр
пищевых систем им. В. М. Горбатова» РАН
Federal State Budgetary Scientific Institution
“V.M. Gorbatov Federal Research Center for
Food Systems of Russian Academy of Sciences”
(Gorbatov Research Center for Food Systems)

Теория и практика переработки мяса
“Teoriä i praktika pererabotki mäsa”
Theory and practice of meat processing
www.meatjournal.ru

Учредитель:

Федеральное государственное
бюджетное научное учреждение
«Федеральный научный центр
пищевых систем
им. В. М. Горбатова» РАН
109316, Россия, Москва,
Талалихина, 26

Издатель:

Федеральное государственное
бюджетное научное учреждение
«Федеральный научный центр
пищевых систем
им. В. М. Горбатова» РАН
109316, Россия, Москва,
Талалихина, 26

Редакция:

Федеральное государственное
бюджетное научное учреждение
«Федеральный научный центр
пищевых систем
им. В. М. Горбатова» РАН
109316, Россия, Москва,
Талалихина, 26
Тел.: +7-495-676-93-51
e-mail: a.zakharov@fncps.ru

Типография:

Федеральное государственное
бюджетное научное учреждение
«Федеральный научный центр
пищевых систем
им. В. М. Горбатова» РАН
109316, Россия, Москва,
Талалихина, 26

Журнал зарегистрирован в Роскомнадзоре

Регистрационные данные:

ПИ № ФС77-71611 от 13.11.2017 года

ЭЛ № ФС77-71609 от 13.11.2017 года

Издается с 2015 года

Контент доступен под лицензией

Creative Commons Attribution 4.0 License

Свободная цена

Периодичность — 4 номера в год

Подписной индекс в каталоге «Пресса России» 38871

Подписано в печать 30.09.2020

Дата выхода из печати 10.10.2020

Тираж 1000 экз. Заказ № 311

© ФНЦПС, 2020

ISSN 2414-438X (Print)

ISSN 2414-441X (Online)

Префикс DOI: 10.21323/2414-438X

Редакционная коллегия:

Главный редактор:

Лисицын Андрей Борисович, доктор технических наук, профессор, Академик
РАН, руководитель научного направления, ФГБНУ «Федеральный научный центр пищевых
систем им. В.М. Горбатова» РАН, Москва, Россия

Заместитель главного редактора:

Чернуха Ирина Михайловна, доктор технических наук, профессор, Академик
РАН, руководитель научного направления, ФГБНУ «Федеральный научный центр
пищевых систем им. В.М. Горбатова» РАН, Москва, Россия

Научные редакторы:

Горлов Иван Федорович, доктор сельскохозяйственных наук, профессор,
академик РАН, научный руководитель, ФГБНУ «Поволжский научно-
исследовательский институт производства и переработки мясомолочной
продукции», Волгоград, Россия

Есимбеков Жанибек Серикбекович, PhD, доцент, кафедра «Технологическое
оборудование и машиностроение», Государственный университет имени
Шакарима, Семей, Республика Казахстан

Galia Zamaratskaia, кандидат технических наук, доцент, научный сотрудник,
Шведский университет аграрных наук, г. Упсала, Швеция

Просекос Александр Юрьевич, доктор технических наук, профессор, член-
корреспондент РАН, ректор ФГБОУ ВО «Кемеровский государственный
университет», Кемерово, Россия

Sakata Ryoichi, Доктор, профессор сельскохозяйственных наук, профессор,
Университет Азабу, Сагамихара, Япония

Tomasevic Igor, кандидат технических наук, доцент, факультет пищевых
технологий и биохимии, кафедра технологии продуктов животного
происхождения, Белградский университет, Белград, Сербия

Горбунова Наталия Анатольевна, кандидат технических наук,
Ученый секретарь, ФГБНУ «Федеральный научный центр пищевых систем
им. В.М. Горбатова» РАН, Москва, Россия

Выпускающий редактор:

Захаров Александр Николаевич, кандидат технических наук,
старший научный сотрудник, заведующий редакционно-издательского отделом,
ФГБНУ «Федеральный научный центр пищевых систем им. В.М. Горбатова» РАН,
Москва, Россия

Члены Редакционной коллегии:

Баженова Баяна Анатольевна, доктор технических наук, профессор,
профессор, кафедра «Технология мясных и консервированных продуктов»
ФГБОУ ВПО Восточно-Сибирский государственный университет технологий
и управления. Улан-Удэ, Россия

Белозеров Георгий Автономович, доктор технических наук,
член-корреспондент РАН, директор, Всероссийский научно-исследовательский
институт холодильной промышленности — филиал Федерального
государственного бюджетного научного учреждения «Федеральный научный
центр пищевых систем им. В.М. Горбатова» РАН, Москва, Россия

Dederer Irina, Кандидат технических наук, научный сотрудник,
Институт Макса Рубнера, Кульмбах, Федеративная Республика Германия
Djordjević Vesna, доктор, директор Института гигиены и технологии мяса,
Белград, Сербия

Дунченко Нина Ивановна, доктор технических наук, профессор,
заведующая кафедрой «Управление качеством и товароведения продукции»,
ФГБОУ ВО «Российский государственный аграрный университет — Московская
сельскохозяйственная академия имени К.А. Тимирязева», Москва, Россия

Кочеткова Алла Алексеевна, доктор технических наук, профессор, руководитель
лаборатории пищевых биотехнологий и специализированных продуктов,
ФГБНУ «Федеральный исследовательский центр питания, биотехнологии
и безопасности пищи», Москва, Россия

Меленцев Алексей Викторович, кандидат экономических наук, директор
Научно-производственного республиканского дочернего унитарного
предприятия «Институт мясо-молочной промышленности» Республиканского
унитарного предприятия «Научно-практический центр Национальной академии
наук Беларуси по продовольствию», Минск, Республика Беларусь

Мирошников Сергей Александрович, доктор биологических наук, профессор,
член-корреспондент РАН, директор, ФГБНУ «Федеральный научный центр
биологических систем и агротехнологий», Оренбург, Россия

Римарева Любовь Вячеславовна, доктор технических наук, профессор,
Академик РАН, главный научный сотрудник, Всероссийский научно-
исследовательский институт пищевой биотехнологии — филиал
ФГБНУ «Федеральный исследовательский центр питания, биотехнологии
и безопасности пищи», Москва, Россия

Рудь Андрей Иванович, доктор сельскохозяйственных наук, главный научный
сотрудник отдела генетики, биотехнологии и технологии в свиноводстве,
ФГБНУ «Федеральный научный центр животноводства — Всероссийский
научно-исследовательский институт животноводства имени академика
Л.К. Эрнста», Подольск, Россия

Семенова Анастасия Артуровна, доктор технических наук, профессор,
заместитель директора, ФГБНУ «Федеральный научный центр пищевых
систем им. В.М. Горбатова» РАН, Москва, Россия

Ханферьян Роман Авакович, доктор медицинских наук, профессор, кафедра
иммунологии и аллергологии, Медицинский институт, Российский университет
дружбы народов, Москва, Россия

**Министерство науки и высшего образования
Российской Федерации
Minister of Science and Higher Education
of the Russian Federation**

Федеральное государственное бюджетное
научное учреждение «Федеральный научный центр
пищевых систем им. В. М. Горбатова» РАН
Federal State Budgetary Scientific Institution
“V.M. Gorbatov Federal Research Center for
Food Systems of Russian Academy of Sciences”
(Gorbatov Research Center for Food Systems)

Теория и практика переработки мяса
“Teoriā i praktika pererabotki mēsa”
Theory and practice of meat processing
www.meatjournal.ru

Учредитель:

Федеральное государственное
бюджетное научное учреждения
«Федеральный научный центр
пищевых систем
им. В. М. Горбатова» РАН
109316, Россия, Москва,
Талалихина, 26

Founder:

Federal State Budgetary Scientific
Institution “V.M. Gorbatov
Federal Research Center
for Food Systems of Russian
Academy of Sciences”
Talalikhina str. 26, Moscow,
Russia, 109316

Издатель:

Федеральное государственное
бюджетное научное учреждения
«Федеральный научный центр
пищевых систем
им. В. М. Горбатова» РАН
109316, Россия, Москва,
Талалихина, 26

Publisher:

Federal State Budgetary Scientific
Institution “V.M. Gorbatov
Federal Research Center
for Food Systems of Russian
Academy of Sciences”
Talalikhina str. 26, Moscow,
Russia, 109316

Редакция:

Федеральное государственное
бюджетное научное учреждения
«Федеральный научный центр
пищевых систем
им. В. М. Горбатова» РАН
109316, Россия, Москва,
Талалихина, 26
Тел.: +7-495-676-93-51
e-mail: a.zakharov@fncps.ru

Editorial Office:

Federal State Budgetary Scientific
Institution “V.M. Gorbatov
Federal Research Center
for Food Systems of Russian
Academy of Sciences”
Talalikhina str. 26, Moscow,
Russia, 109316
Tel.: +7-495-676-93-51
e-mail: a.zakharov@fncps.ru

Типография:

Федеральное государственное
бюджетное научное учреждения
«Федеральный научный центр
пищевых систем
им. В. М. Горбатова» РАН
109316, Россия, Москва,
Талалихина, 26

Printing Office:

Federal State Budgetary Scientific
Institution “V.M. Gorbatov
Federal Research Center
for Food Systems of Russian
Academy of Sciences”
Talalikhina str. 26, Moscow,
Russia, 109316

Журнал зарегистрирован в Роскомнадзоре

Регистрационные данные:

ПИ № ФС77-71611 от 13.11.2017 года

ЭЛ № ФС77-71609 от 13.11.2017 года

Published in 2015

This work is licensed under a

Creative Commons Attribution 4.0 License

Free price

Frequency — 4 issues a year

Subscription index in the catalogue “Press of Russia” 38871

Signed print 30.09.2020

Released from press 10.10.2020

Circulation — 1000 copies. Order № 311

© FNCPS, 2020

ISSN 2414-438X (Print)

ISSN 2414-441X (Online)

DOI-prefix: 10.21323/2414-438X

Editorial board:**Editor-in-Chief:**

Andrey B. Lisitsyn, Doctor of technical sciences, Professor, Academician of RAS, Scientific supervisor, V.M. Gorbatov Federal Research Center for Food Systems of Russian Academy of Sciences, Moscow, Russia

Deputy Editor-in-Chief:

Irina M. Chernukha, Doctor of technical sciences, professor, Academician of RAS, Head of the scientific direction, V.M. Gorbatov Federal Research Center for Food Systems of Russian Academy of Sciences, Moscow, Russia

Scientific Editors:

Ivan F. Gorlov, Doctor of agricultural sciences, professor, Academician of RAS, Scientific supervisor, Povolzhskiy Research Institute of Production and Processing of Meat and Dairy Products, Volgograd, Russia

Zhanibek S. Yessimbekov, PhD, docent, Department of Mechanical Engineering and Mechanics, Shakarim State University, Semey, Republic of Kazakhstan

Galia Zamaratskaia, Candidate of technical sciences, docent, Research worker, Swedish University of Agricultural Sciences, Uppsala, Sweden

Aleksandr Yu. Prosekov, Doctor of technical sciences, professor, corresponding members of RAS, Rector, Kemerovo State University, Kemerovo, Russia
Sakata Ryoichi, PhD, doctor, professor of agricultural sciences, Azabu University, Sagami-hara, Japan

Tomasevic Igor, PhD, Associate Professor, Animal Source Food Technology Department, Faculty of Agriculture, University of Belgrade, Belgrade, Serbia

Natalia A. Gorbunova, Candidate of technical sciences, Academic Secretary, V.M. Gorbatov Federal Research Center for Food Systems of Russian Academy of Sciences, Moscow, Russia

Production editor:

Aleksandr N. Zakharov, candidate of technical sciences, senior research worker, Head or research worker, Head of the Department of Editorial and Publishing, V.M. Gorbatov Federal Research Center for Food Systems of Russian Academy of Sciences, Moscow, Russia

Members of the editorial board:

Baiana A. Bazhenova, Doctor of technical sciences, professor, Professor, chair “Meat and canned product technology”, East Siberia State University of Technology and Management, Ulan-Ude, Russia

Georgy A. Belozherov, Doctor of technical sciences, corresponding members of RAS, Director, All-Russian Scientific Research Institute of Refrigeration Industry — Branch of V.M. Gorbatov Federal Research Center for Food Systems of RAS, Moscow, Russia

Irina Dederer, Candidate of technical sciences, Research worker, Max Rubner-Institut, Kulmbach, Germany

Vesna Djordjevic, Doctor, Director, the Institute of Meat Hygiene and Technology, Belgrad, Serbia

Nina I. Dunchenko, Doctor of technical sciences, professor, the Head of the chair “Product quality management and merchandise knowledge”, Russian State Agrarian University — Moscow Timiryazev Agricultural Academy, Moscow, Russia

Alla A. Kochetkova, Doctor of technical sciences, professor, the Head of the “Laboratory of food biotechnologies and specialized products”, Federal Research Centre of nutrition, biotechnology and food safety, Moscow, Russia

Aliaksei V. Meliashchenia, Candidate of economical sciences, Director, Institute of Meat and Dairy Industry of the Republican Unitary Enterprise “The Scientific-practical Center of the National Academy of Sciences of Belarus for food”, Minsk, The Republic of Belarus

Sergey A. Miroshnikov, Doctor of biological sciences, professor, corresponding member of RAS, Director, Federal Research Centre of Biological Systems and Agrotechnologies RAS, Orenburg, Russia

Liubov V. Rimareva, Doctor of technical sciences, professor, Academician of RAS, Leading scientific worker, All-Russian Scientific Research Institute of Food Biotechnology — branch Federal Research Centre of nutrition, biotechnology and food safety, Moscow, Russia

Andrey I. Rud, Doctor of agricultural sciences, Chief research worker of the Department of Genetics, biotechnology and technology in pig, L.K. Ernst Federal Science Center for Animal Husbandry, Podolsk, Russia

Anastasiya A. Semenova, Doctor of technical sciences, professor, Deputy Director, V.M. Gorbatov Federal Research Center for Food Systems of Russian Academy of Sciences, Moscow, Russia

Roman A. Khanferyan, Doctor of medical sciences, Professor, Department of Immunology and Allergology, Medical Institute, Peoples’ Friendship University of Russia, Moscow, Russia

CONTENTS

Elizaveta V. Kryuchenko, Yulya A. Kuzlyakina, Valentina S. Zamula, Irina M. Chernukha ALLERGENOMICS AND ANALYSIS OF CAUSES OF UNINTENTIONAL INCORPORATION OF SUBSTANCES CAPABLE OF CAUSING IGE-MEDIATED FOOD ALLERGY INTO MEAT PRODUCTS	4
Andrey B. Lisitsyn, Irina M. Chernukha, Marina A. Nikitina A SYSTEM APPROACH TO SIMULATION OF INDIVIDUAL FOOD PRODUCTS	12
Ekaterina R. Vasilevskaya, Anastasiya G. Akhremko, Ekaterina K. Polishchuk, Liliya V. Fedulova STUDY OF THE FUNCTIONAL PRODUCT'S PROTEIN COMPOUNDS DIGESTION FEATURES.....	18
Isabella L. Stefanova, Liudmila V. Shakhnazarova, Anastasia Yu. Klimenkova, Inna M. Sorokina THE CHANGES IN THE AMINO AND FATTY ACID PROFILES IN THE SEMIFINISHED FOODSTUFFS BASED ON BROILER MEAT AND COMPONENTS OF CHICKEN EGGS AFTER DIFFERENT TYPES OF THERMAL TREATMENT	22
Konstantin A. Kurbakov, Valentina N. Zhulinkova, Mihail Yu. Minaev DEVELOPMENT OF HIGH SENSITIVE REAL-TIME PCR TO DETECT MUSTARD AND OTHER ALLERGENS OF THE FAMILY <i>BRASSICACEAE</i> IN FOOD SAMPLES	28

ALLERGENOMICS AND ANALYSIS OF CAUSES OF UNINTENTIONAL INCORPORATION OF SUBSTANCES CAPABLE OF CAUSING IGE-MEDIATED FOOD ALLERGY INTO MEAT PRODUCTS

Elizaveta V. Kryuchenko, Yulya A. Kuzlyakina*, Valentina S. Zamula, Irina M. Chernukha

V. M. Gorbatov Federal Research Center for Food Systems of Russian Academy of Sciences, Moscow, Russia

Keywords: food allergy, allergens, causes, unintentional ingestion, meat products

Abstract

The article discusses the definition and mechanism of IgE-mediated food allergy, provides an overview of the legal regulation of the production and labeling of allergen-containing food products. In order to prevent the inadvertent appearance of allergens in products during their production, an allergenomics procedure is required — a comprehensive assessment of the allergic potential of a food product: allergenicity of product ingredients, risk analysis, and the procedure for managing allergens in the production.

Introduction

Allergic diseases have gained an enormous scale in the world, both in developed and developing countries. According to the data of the World Health Organization, a number of patients with allergies have increased in Russia by 20% over the last decade. According to scientists' forecasts, this number will grow as a majority of factors causing allergic reactions are linked with modern lifestyle. As the growth in the number of allergic reactions associated with food consumption is rapid, a food allergy is necessary to regard as one of the main public health problems [1,2,3,4]. The size of population with food allergies is different in various countries. For example, the prevalence of food allergies is 4.6% in Spanish population and 19.1% in Australian population [5]. According to various data, the prevalence of food allergies in the Russian Federation is 30% to 56% in children [6] and about 20% in adults suffering from atopic dermatitis [7].

At present, there are no available methods for prevention or treatment of food allergies: the only method to maintain remission in a patient is to exclude intake of a food allergen and treatment in the case of exacerbation includes only managing symptoms as they are revealed [9,10]. Individuals suffering from food allergies have to adhere to special diets to avoid allergic reactions [11]. Governments of many European countries acknowledged an importance of problems associated with food allergies and set requirements on the legislative level for managing allergens and labeling products containing allergens [12]. Nevertheless, the control of allergens in a processing enterprise and throughout the ingredient supply chain is a complex task for producers in the conditions of globalized economy [12]. When ingredients are obtained from suppliers of different regions and foreign suppliers, the risk of increasing a likelihood of unintentional presence of allergens in food products appears, which can lead to potential threats to food safety as well as economic threats, which is evident from large scale recalls affected many food companies [13,14,15,16].

Technologists and other specialists directly working with food products should have insight into forms of allergic diseases and their causes, as well as the causes of unintentional incorporation of allergens into products upon their production. It will be interesting for specialists of meat processing plants that, as a rule, meat per se does not cause allergic reactions, which, unfortunately, cannot be said about additives that are used in meat product manufacture [17].

At the same time, it is necessary to consider socio-psychological consequences of the food allergy risk, in particular, factors determining quality of life [18].

Main part

1. Definition and mechanism of IgE-mediated food allergy

A food allergy is an immune response to the contact of the body with food. A reaction can be mediated by IgE release, activation of T-cells or tissue basophils [19].

The IgE type I reactions are distinguished by recognition of IgE specific epitopes (linear or conformational) within a soluble antigen to trigger mast cell activation [20,21,22]. As a rule, the IgE-mediated food allergy is characterized by rapid onset: skin (urticarial, Quincke's edema, exacerbation of atopic dermatitis/eczema), gastrointestinal (nausea, vomiting, diarrhea) and/or respiratory symptoms [23,24] appear in patients usually in an interval from several minutes to 2 hours.

The mechanism involved in the IgE-mediated food allergic reactions is shown in Figure 1. There are two stages in the development of the IgE-mediated food allergy: the sensitization phase and manifestation phase [24]. Sensitization can occur at any age and does not always emerge at the first allergen exposure. Sensitization does not have symptoms and consists of adsorption, processing and presentation of an allergen, activation of T-cells and B-cells, development of oral tolerance or allergic sensitivity and synthesis of antigen-specific IgE-antibodies by plasma cells. Allergen-specific IgE

FOR CITATION:

Chernukha I. M., Kuzlyakina Yu.A., Zamula V. S., Kryuchenko E. V. Allergenomics and analysis of causes of unintentional incorporation of substances capable of causing IgE-mediated food allergy into meat products. *Theory and practice of meat processing*. 2020;5(3): 4–11. <https://doi.org/10.21323/2414-438X-2020-5-3-4-11>

binds to the surface of mast cells in different connective tissues (the gastrointestinal system, respiratory tract, skin) and basophils in blood. Cross-linking of allergens with IgE on the surface of mast cells or basophilic membrane triggers release of histamine and other chemotactic mediators responsible for clinical allergic symptoms. Histamine, prostaglandins and leukotrienes can promote contraction of smooth tissue in the blood vessels, gastrointestinal tract and respiratory tract and increase permeability and dilation of vessels, increase mucus secretion and increase chemotaxis of eosinophils, neutrophils and mononuclear cells. Mediators are released into the blood flow and can cause systemic reactions affecting several tissues and organs [24,25].

In addition to IgE-mediated food allergy, which symptoms are known and the mechanism is described, there is non-IgE-mediated food allergy, which pathogenesis is not fully understood.

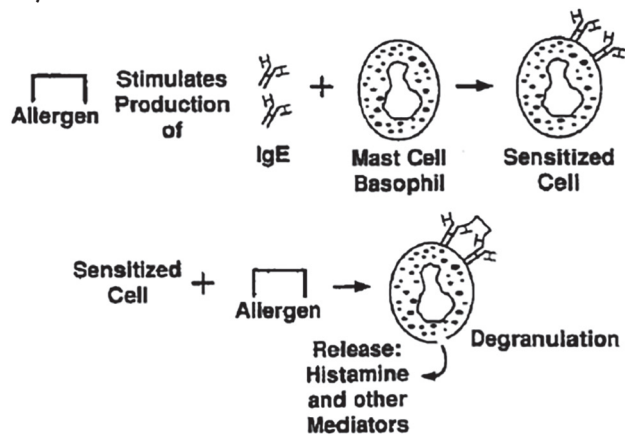


Figure 1. Mechanism of IgE-mediated food allergy

Almost all products with natural proteins can cause allergic reactions in certain individuals. However, an impact of food proteins not always leads to the production of protein-specific IgE antibodies and only a small percent of food proteins was identified as allergens [26]. A majority of food allergens are water- or salt-soluble glycoproteins with acidic isoelectric end points that are comparatively resistant to processing, food preparation, proteolysis and digestion processes [27].

2. Prevalence of IgE-mediated food allergy

As mentioned above, food allergy is a problem of public health worldwide as 5–10% of children and 3–4% of adults suffer in the western countries according to medical estimates. Awareness about food allergies is growing and up to 35% of people self-diagnose food allergies. The prevalence of food allergies is increasing in the world, although specific causes have not been revealed [28,29,30].

There are many theories about growing prevalence of food allergies: the genetic factors, number of caesarean sections, hygienic hypothesis, time and route of the first contact with food allergens, changes in nutrition habits, food processing and levels of vitamin D exposure [30]. No large-scale changes in population genetics can explain the rise in food allergies [31]. Epigenetics is the study of heritable and non-inherited

changes in the gene function, which occur without changes in the DNA nucleotide sequence. Epigenetic changes caused by changes in a diet and environmental impact were associated with the development of asthma and allergic rhinitis, but not with food allergy [32,33]. The hygienic hypothesis assumes that an increase in the infection level at an early age has a protective action on the development of allergies, asthma and other atopic diseases [34,35,36,37]. The hypothesis about the double action of an allergen states that tolerance emerges due to peroral food exposure and allergic sensitization due to skin exposure [38]. Inflammation caused by eczema reduces the effectiveness of epidermal barrier protein and opens a possibility of allergen protein impact and production of food allergen-specific T-cells in unprotected skin [39]. Low levels of peanut are accessible to infants in household conditions after cleaning providing skin exposure for individuals at risk [40]. The time of peanut introduction into a diet influenced significantly the prevalence of peanut allergy among Israeli schoolchildren [41]. Israeli children consumed more peanut during the first year of life compared to UK children and the prevalence of peanut allergy was 0.17% in Israel and 1.85% in the UK. Changes in atopy, social class or genetic background did not have a significant effect [41]. In the USA, the number of people with peanut allergy doubled over four years (2006–2010). With that, the incidence of anaphylactic shock caused by peanut doubled over the five-year period [42]. In addition, the form of peanut consumption can determine the appearance of allergic reaction. Stability and allergenicity of allergenic proteins can be altered upon food processing. For example, peanut roasting affects the stability of peanut allergens via the Maillard reaction and modified peanut allergens have an increased ability to bind IgE [43,44]. Nevertheless, there is no reliable evidence that allows linking changes in nutrition habits or food industry with the rise in the food allergy prevalence [41].

In the USA, milk, eggs and peanut are the most common allergenic food among children, while adults more often suffer from allergies to shellfish, peanut and tree nuts [30]. Many children will outgrow food allergies and become more tolerant to milk, eggs, soybean and wheat. Allergies to peanut, and tree nuts and shellfish seldom reduce with age [30,45,46]. Allergies to milk and eggs are prevalent worldwide; however, other main food allergens will vary among regions depending on cultural and dietetic habits [47]. Food allergies are potentially dangerous for life. Once ingested, food allergens can cause anaphylactic shock and human death. Using the developed method for studying possible cases of fatal anaphylaxis by the tryptase level from mast cells and allergen-specific antibodies to immunoglobulin E (IgE) in serum of people died from anaphylactic shock, John W. Yunginger et al. [48] established increased serum tryptase levels (12 ng/ml to 150 µg/ml) in nine of nine fatal cases caused by food. Serum IgE antibodies were increased in eight of eight studied fatal cases due to the food allergic reaction. According to the medical statistics data, 100 to 200 deaths from anaphylactic shock due to food allergies are recorded in the USA each year [9].

Catering establishments and educational institutions remain to be the most common places of fatal allergic reactions, and peanut accounts for more than 50% of deaths linked with food allergies in the USA [49].

The problem of the allergic response to innovative food additives and products with them deserves close attention. These are GMO food ingredients. Studies show that albumin, globulin, gluten of transgenic wheat varieties can cause asthma and IgE food allergy [50]. It is also of interest to assess an allergenicity risk of protein from non-traditional sources such as insects.

3. Legislative requirements

Initially, the list of top priority allergens was published in Codex Alimentarius in 1999. Later on, this list became a starting point for the European Commission and other state organizations to publish the special legislative act that regulates labeling of food products containing the ingredients from the list [51].

In foreign countries, legislative requirements that included a list of allergens and the processes of their control were developed:

- The Regulation (EU) no 1169/2011 on the provision of food information to consumers. The Annex II of this document lists 14 groups of food products that cause allergies, which should be mandatory on a product label if they are used as ingredients irrespective of their quantity [52].
- Directive 2003/89/EC as regards indication of the ingredients present in foodstuffs (European Union)
- Directive 2005/26/EC on allergens (European Union)
- Federal Legislation. Section 201–210 (USA)
- Food Allergen Labeling and Consumer Protection Act of 2004
- Australia New Zealand Food Standards Code — Standard 1.2.3 [2].

Recommendation 24–2017 of the Scientific Committee of the Federal Agency for the Safety of the Food Chain (FASFC) (Belgium) regarding reference doses of allergens listed in Annex II of Regulation (EC) No. 1169/2011 of October 25, 2011.

In November 2015, USDA's Food Safety and Inspection Service (FSIS) issued guidelines to assist producers of meat, poultry and processed egg products in attempt to reduce side reactions to food allergens. This guidance includes measures for prevention and control of potentially allergenic ingredients, packaging, labeling, control lists and training [53].

In 2018, the Proposed Draft Code of Practice on Food Allergen Management for Food Business Operators was placed on the official site of the Food and Agriculture Organization of the United Nations (FAO) for public discussion; it was planned to consider the possibility of its adoption at the 43th session of the Codex Alimentarius Commission on July 6–11 2020 in Rome (Italy).

In Russia, the list of most common food allergens, the consumption of which may cause allergic reactions or is contraindicated in certain types of diseases, is given in the

Technical Regulation of the Customs Union “Food products in part of their labeling” (TR CU022/2011); it is fully harmonized with the EC legislation and contains the following products:

- 1) peanut and products of its processing;
- 2) aspartame and aspartame-acesulfame salt;
- 3) mustard and products of its processing;
- 4) sulphur dioxide and sulphites if their total content exceeds 10 milligrams per 1 kilogram or 10 milligrams per one liter in terms of sulphur dioxide;
- 5) cereals, containing gluten and products of their processing;
- 6) sesame and products of its processing;
- 7) lupin and products of its processing;
- 8) molluscs and products of their processing;
- 9) milk and products of its processing (including lactose);
- 10) nuts and products of their processing;
- 11) crustaceans and products of their processing;
- 12) fish and products of its processing (excluding fish gelatin used as a basis in preparations containing vitamins and carotenoids);
- 13) celery and products of its processing;
- 14) soya and products of its processing;
- 15) eggs and products of its processing [54].

In addition, according to the requirements of TR CU022/2011 “Food products in part of their labeling”, components capable of causing allergic reactions are indicated in the food product composition irrespective of their quantity. These measures are aimed at ensuring provision of timely information to consumers with food allergies for correct composition of their diets.

4. Causes of unintentional incorporation of allergens into meat products upon its production

When a problem with meat product safety linked with incorrect handling of allergenic ingredients arises, not only consumer health but also reputation and economic stability of meat industry enterprises are threatened. To avoid realization of such risks, specialists of meat processing enterprises should analyze causes of unintentional incorporation of allergens into meat products upon their production. It is necessary to assess every technological stage supporting the process where realization of the risk of the undeclared allergen presence in meat products is possible, and analyze information related to unintentional incorporation of allergens into products during their processing [23,55,56,57, 58, 59].

As a result of the analysis of the likelihood of allergen realization, the following causes of unintentional incorporation of allergens into meat products can be identified (Table 1).

Activities on meat product manufacture are different by their character and not all causes of allergen incorporation into meat products highlighted in table 1 are applicable to a particular enterprise or process. The common denominator of all examined causes is a requirement for hazard analysis and absence of information on realization of this requirement [60].

Table 1. Causes of unintentional incorporation of allergens into meat products

Life cycle processes, supporting processes, management processes	Causes of unintentional incorporation of allergens
Purchase of raw materials, specifications (incoming control)	<ul style="list-style-type: none"> — absence of procedures for assessing suppliers; — accompanying documents are not analyzed upon raw material entrance for obtaining corresponding information about an allergen or any changes; — absence of information on a passport of every pallet/box/bag about the presence of an allergen (enterprises can use color coding, labeling or other means for identification of allergenic ingredients); — mishandling of damaged containers, boxes, bags with allergens, which leads to cross-contamination upon receiving; — absence of information about GMO ingredients; — absence of information about non-traditional protein sources
Planning of production	<ul style="list-style-type: none"> — joint storage and transfer across an enterprise of ingredients containing allergens and ingredients free from allergens; — nonuse of clear designation for separation zones of intermediate storage and transfer; — absence of physical barriers; — nonuse of special trays, containers, appliances; — allergenic ingredients are not identified by labeling or color coding; — closed containers are not used; — procedures for cleaning from spillage or damaged containers with allergens are not used and documented; — succession of manufacturing allergenic products after allergen free products is not planned; — risk of allergenic dust migration during processing is not assessed; — absence of control for reincorporation of a product into a process
Sanitary	<ul style="list-style-type: none"> — time schedule for sanitary treatment is not made; — absence of instructions for equipment cleaning; — absence of equipment cleaning immediately after manufacturing food products with allergens; — cleaning quality is not controlled; — absence of allocated tools for cleaning; — absence of documented rules for cleaning of spillage; — absence of documented rules for disassembly of equipment when cleaning
Training and instruction of personnel	<ul style="list-style-type: none"> — absence of training on allergen awareness and control of personnel according to their job responsibilities; — hand washing is not controlled; — special clothing is not allocated and its timely exchange is not controlled; — control of product rework is absent; — waste control is absent; — control of appliances use is absent
Packaging and labeling	<ul style="list-style-type: none"> — absence of packaging control; — change of packaging lines is not controlled; — high level of problems with mislabeling
Food safety system	<ul style="list-style-type: none"> — one of hazard types (allergen) was not considered when analyzing a likelihood of hazard factor realization and severity of its consequences; — failure to support decisions made in the course of hazard analysis; — failure to assess a likelihood of hazard factor realization and severity of its consequences; — failure to implement effectively control means to support decisions made in the course of hazard analysis

To minimize possible unintentional incorporation of allergens into food products, food industry enterprises develop and introduce a complex of measures within a framework of allergen management programs [55].

Advanced companies that have been working for many years according to the international food safety and quality standards determined years ago the ways of the development with regard to designing allergen management programs. As for small and medium-sized businesses, the scantiness of information resources in the sphere of implementation tools does not allow them to compete with giant manufacturers [61].

Introduction of allergen management should be regarded as an extension of the existing system of food safety management.

At the initial stage of work in this direction, it is expedient to analyze the following factors:

- total quantity of allergens that can provoke a reaction in sensitive people (these data are relative as different people can have different levels of sensitivity and sensitivity of a person can vary under different circumstances);
- how frequent population consuming allergen-containing food products has general adverse reactions;
- whether there are any subgroups of the population that are in the special risk zone (infants and children). These subgroups include people who restrict food choices due to diets;
- relative allergenicity of the component being used; moreover, if a product was processed, a corresponding protein can be absent and, therefore, it will not present a risk of cross-contamination with an allergen;
- origin of particular ingredients, their geographic and manufacturing environment [57].

Then, a likelihood of cross-contamination at each stage of food production process is assessed beginning from the incoming control of food raw materials to finished product sale. With that, it is necessary to assess a physical form of used allergens, for example, a liquid and powder present a different degree of the cross-contamination risk. For instance, during powder milk weighing, it can be introduced into a product through a ventilation system or from personnel clothes; while introduction of liquid milk is less likely when adhering to certain measures (separation with physical barriers, a distance between products).

When an unacceptable contamination risk was identified, it is necessary to apply measures aimed at reduction of unintentional presence of allergens in a product to the fullest extent possible. To this end, Good Manufacturing Practice (GMP) has been successfully used in the framework of production process organization. To ensure food safety, GMP requires maintenance of strict discipline by all personnel [63]. Key aspects of allergen management in meat product manufacture are presented in Figure 2.



Figure 2. The main areas for consideration when creating allergen management system

A manufacturer should know about the presence of allergens in all used raw materials, which is achieved when working with suppliers and upon incoming control of accompanying documents on raw materials. A manufacturer should request information from suppliers regarding the content of food allergens in raw materials in a form of:

- the main components indicated in the composition (for example, soy plant protein in the composition of the complex food additive);
- auxiliary components (for example, a food additive produced from an allergenic source, for example, amylase from wheat);
- undeclared components introduced due to cross-contamination with an allergen upon production.

Raw material suppliers should be aware about risks that may realize as a result of product contamination with aller-

gens and provide corresponding information. Components should be fully described on a label and in specifications; the use of generalized names of used ingredients such as “plant oils and fats” is unacceptable [53,56].

When placing in a manufacturer’s warehouse after incoming control, raw materials containing allergens should be identified; it is expedient to provide separate storage of such ingredients.

The only approach to full exclusion of cross-contamination with allergens during production process is the use of separate production areas; however, it is often impossible. There are other measures for separating products with allergens from products without allergens:

- separation of production into zones; establishment of physical barriers between production lines;
- provision of allocated equipment, appliances and containers;
- minimization of unnecessary material movement; proper planning of production cycles including equipment cleaning between production cycles;
- organization of individual air supply where it is possible, and so on.

At the stage of incoming control of the main raw materials and auxiliary materials, they are checked on correspondence to normative technical documentation including information about the presence of allergens. Training of responsible employees on allergen awareness and their control according to job responsibilities are carried out. The control of corresponding documents, identification of incoming raw materials and other materials regarding correspondence to information, visual assessment are performed. Then, clear labeling is carried out indicating whether there is a potential allergen (enterprises can use color coding or other means for identification of allergenic ingredients) and batches of incoming raw materials and other materials are placed separately [62]. At the planning stage, it is necessary to segregate zones for storage, production of the main raw materials, auxiliary materials with and without allergens; however, if there is no such possibility, other methods are used. Zones for allergen storage are prepared and established. Special transport containers (marked or color coded) are used. Allergenic raw materials are placed in an allocated and marked warehouse zone separately from raw materials without allergens; physical barriers are used. Instructions have been developed with regard to prevention of cross contamination and are distributed in necessary locations [58, 59]. When transporting allergenic raw materials from a warehouse to the area of spices composition, special marked closed containers are used.

Routes for transportation of allergens and non-allergens, finished products and waste are separated by time (space) to prevent cross-contamination. After transportation, facilities are cleaned along the route of transportation and sanitary treatment of transport equipment is applied. When storing and using allergens, racks, weighing scales, appliances (shovels, small containers, bag), places for storage

of cleaning appliances and cleaning appliances per se are marked. Personnel use special clothes; the control of its timely exchange is performed. Operation of a ventilation system is controlled. In meat product manufacture, production of allergenic products after products that are free from allergens is planned. After the end of manufacturing process, equipment and appliances are thoroughly cleaned. It is necessary to make schedules of sanitary treatment and instructions, control quality of equipment cleaning, allocate tools, develop rules for cleaning of spillage and disassembly of equipment upon cleaning. Also, it is necessary to carry out identical measures and control upon packaging products with allergens and products free from them. All allergenic ingredients should be indicated on a label, product labeling is carried out according to TR CU022/2011[54]. It is necessary to control secondary processing and utilization of food waste. It is stated in article 10 clause 2 of Technical Regulation of the Customs Union 021/2011 "On safety of food products" that a manufacturer should develop, introduce and maintain procedures based on the HACCP principles. In the system framework, it is necessary to analyze risks of the likelihood of hazard factor realization and severity of its consequences. Previously, three hazard types (biological, chemical, physical) were examined; now allergens are also analyzed.

At present, specialists of the V. M. Gorbatov Federal Research Center for Food Systems of Russian Academy of Sciences have been developing a draft of GOST R "Meat industry. Order of development of allergen management program for meat industry". The present standard gives recommendations to producers on the development of procedures for determination of allergens in the process of production as well as on realization of measures for allergen management including control measures for:

- managing a level of a hazard for meat product safety, which is characteristic for a product and environment where it is produced;
- managing a likelihood that a production environment will become a source of emergence of hazards for food safety;

— assurance of correct labeling of allergens for packaged finished products.

Meat industry enterprises have a big responsibility in product manufacture regarding the correspondence to the requirements of the legislation and regarding consumers' health. In this connection, it is necessary to develop, introduce and maintain a program for allergen management, analyze the causes of allergen realization and organize resource management to minimize unintentional incorporation of allergens into finished products.

Conclusion

Food allergy is a developing problem of public health, which can have a serious consequences for health of consumers sensitive to food allergens and can even lead to death. There are IgE and IgG food allergies. IgE food allergy is an acute reaction that occurs in 2–3% of population. Any food product that is considered allergenic can cause this type of allergy. About 20% of world population have IgG food allergy. It is characterized by the delayed allergic reaction with lower risk of a severe disease or death. As today the only method to stop a food allergy is complete exclusion of an ingredient that causes an allergy from a diet, the food industry, in particular, its meat branch, has to provide a consumer with reliable information on a product label, as well as exclude unintentional incorporation of allergens into products upon their manufacture. When a threat for meat product safety linked with mishandling of allergenic ingredients arises, specialists of meat processing enterprises should analyze causes of unintentional incorporation of allergens into meat products upon their manufacture. In analysis, it is necessary to assess every technological stage supporting a process, where the realization of the risk of the presence of undeclared allergens in meat products is possible, and analyze information linked with causes of unintentional incorporation of allergens into products upon their manufacture. To control allergens, modern analytical methods such as mass spectrometry are necessary. It is necessary to develop databases of protein sequences to simplify identification of allergenic protein in proteomic investigations.

REFERENCES

1. Karataeva, N.A., Ksrstaeva, L.A., Yusupova, O.I., Inoyanova, Sh. Sh. (2015). Food allergy (review). *Scientific research*, 1, 126–130. (in Russian)
2. Lisitsyn, A.B., Chernukha, I.M., Lunina, O.I. (2017). Food hypersensitivity and products of animal origin resources. *Theory and practice of meat processing*, 2(2), 23–36. <https://doi.org/10.21323/2414-438X-2017-2-2-23-36> (in Russian)
3. Pawankar, R., Canonica, G. W., Holgate, S.H., Lockey, R.F. (2011). WAO White Book on Allergy 2011–2012: Executive Summary. World Allergy Organization. —24 p.
4. Taylor, S.L., Lemanske, R.F., Bush, R.K., Busse, W.W. (1987). Food allergens: structure and immunologic properties. *Annals of Allergy*, 59(5 PART II), 93–99.
5. Crespo, J.F., Rodriguez, J. (2003). Food allergy in adulthood. *Allergy: European Journal of Allergy and Clinical Immunology*, 58(1), 98–113. <https://doi.org/10.1034/j.1398-9995.2003.02170.x>
6. Tihkon, N.M., Vasilevskaya, O.A., Kazanovich, V.V. (2016). Allergic diseases in children: modern aspects. *Proceeding of the 19th Congress of pediatricians of Russia with international participation "Topical problems of pediatrics"*, 294. (in Russian)
7. Simonova, A.V., Kosheleva, I.V., Shadyzheva, L.I. (2016). Optimization of treatment and prevention of exacerbation of atopic dermatitis with account for the main pathological factors. *Therapist*, 5, 67–69. (in Russian)
8. Khaitov, R.M. (2014). Allergology and immunology: national guidance. Moscow: GEOTAR-Media. — 656 p. ISBN 978-5-9704-0903-9. (in Russian)
9. Boyce, J., Assa'ad, Burks, A (2010). Guidelines for the diagnosis and management of food allergy in the United States: report of the NIAID-sponsored expert panel. *Journal of Allergy and Clinical Immunology*, 126(6 SUPPL.), S1-S58.
10. On, P.Y. (2008). Are allergy advisory statements helpful to food allergy patients? *Immunol. Journal of Allergy and Clinical Immunology*, 121(2), 536–537. <https://doi.org/10.1016/j.jaci.2007.11.006>
11. Remington, B.C., Westerhout, J., Meima, M.Y., Blom, W.M., Kruizinga, A.G., Wheeler, M.W., Taylor, S.L., Houben, G.F., Bau-

- mert, J.L. (2020). Updated population minimal eliciting dose distributions for use in risk assessment of 14 priority food allergens. *Food and Chemical Toxicology*, 111259. <https://doi.org/10.1016/j.fct.2020.111259>
12. Yeung, J., Robert, M.-C. (2018). Challenges and Path Forward on Mandatory Allergen Labeling and Voluntary Precautionary Allergen Labeling for a Global Company. *Journal of AOAC International*, 101(1), 70–76. <https://doi.org/10.5740/jaoacint.17-0391>
13. Garber, E.A.E., Parker, C.H., Handy, S.M., Cho, C.Y., Panda, R., Samadpour, M., Reynaud, D.H., Ziobro, G.C. (2016). Presence of Undeclared Food Allergens in Cumin: The Need for Multiplex Methods. *Journal of Agricultural and Food Chemistry*, 65(5), 1202–1211. <https://doi.org/10.1021/acs.jafc.5b05497>
14. Gendel, S. M. (2017). Food Allergen Recalls: The Past as Prologue. Book Chapter in Food Allergens, 95–102. https://doi.org/10.1007/978-3-319-66586-3_5
15. Sayers, R. L., Gethings, L., Wallace, A., Semic-Jusufagic, A., Simpson, A., Barran, P., Gilbert, J., Senyuva, H., Rodgers, A., Bromley, M., Walker, M., Brown, H., Mills, E. N. C. (2016). How Much of a Problem Is Peanut in Ground Cumin for Individuals with Peanut Allergy? *Journal of Allergy and Clinical Immunology*, 137(2), AB142. <https://doi.org/10.1016/j.jaci.2015.12.597>
16. Walker, M. J., Burns, D. T., Elliott, C. T., Gowland, M. H., Mills, E. N. C. (2016). Is food allergen analysis flawed? Health and supply chain risks and a proposed framework to address urgent analytical needs. *The Analyst*, 141(1), 24–35. <https://doi.org/10.1039/c5an01457c>
17. Davydova, R. (2013). Intolerance of food products and labeling of allergens in the EC countries. *Meat branch*, 3(123), 34–37. (in Russian)
18. Voorheis, P., Bell, S., Cornelsen, L., Quaife, M., Logan, K., Marrs, T. et al. (2019). Challenges experienced with early introduction and sustained consumption of allergenic foods in the Enquiring About Tolerance (EAT) study: A qualitative analysis. *Journal of Allergy and Clinical Immunology*, 144(6), 1615–1623. <https://doi.org/10.1016/j.jaci.2019.09.004>
19. Luss, L.V. (2012). Food allergy and food intolerance, terminology, classification, problems of diagnosis and therapy. *Effective Pharmacotherapy*, 7, 48–56. (in Russian)
20. Murphy, K.P., Travers, P., Walport, M. (2008). Janeway's immunobiology. 7th ed., New York: Taylor & Francis Group. ISBN 978-0-8153-4123-9
21. Pomes, A. (2010). Relevant B cell epitopes in allergic disease. *International Archives of Allergy and Immunology*, 152(1), 1–11. <https://doi.org/10.1159/000260078>
22. Untersmayr, E., Jensen-Jarolim, E. (2006). Mechanisms of type I food allergy. *Pharmacology and Therapeutics*, 112(3), 787–798. <https://doi.org/10.1016/j.pharmthera.2006.06.004>
23. Food Allergen Labeling and Consumer Protection Act of 2004 (FALCPA) [Electronic resource: <https://www.fda.gov/food/food-allergens/gluten-free-guidance-documents/regulatory-information/food-allergen-labeling-and-consumer-protection-act-2004-falcpa> Access date 21.04.2020]
24. Taylor, S.L., Hefle, S.L., Bindslev-Jensen, C., Atkins, F.M., André, C., Bruijnzeel-Koomen, C., Burks, A.W., Bush, R.K., Ebisawa, M., Eigenmann, P.A., Host, A., Hourihane, J.O.B., Isolauri, E., Hill, D.J., Knulst, A., Lack, G., Sampson, H.A., Moneret-Vautrin, D.A., Rancé, F., Vadas, P.A., Yunginger, J.W., Zeiger, R.S., Salminen, J.W., Madsen, C., Abbott, P., (2004). A consensus protocol for the determination of the threshold doses for allergenic foods: how much is too much? *Clinical Experimental Allergy*, 34(5), 689–695. <https://doi.org/10.1111/j.1365-2222.2004.1886.x>
25. Fraser, O., Sumar, S., Jones, M., Sumar, N. (2001). Mechanisms in adverse reactions to foods: a review. *Nutrition & Food Science*, 31(4), 170–178. <https://doi.org/10.1108/00346650110392235>
26. Hefle, S.L., Furlong, T.J., Niemann, L., Lemon-Mule, H., Sicherer, S., Taylor, S.L. (2007). Consumer attitudes and risks associated with packaged foods having advisory labeling regarding the presence of peanuts. *Journal of Allergy and Clinical Immunology*, 120(1), 171–176. <https://doi.org/10.1016/j.jaci.2007.04.013>
27. Taylor, S.L., Lehrer, S.B. (1996). Principles and characteristics of food allergens. *Critical Reviews in Food Science and Nutrition*, 36(sup001), 91–118. <https://doi.org/10.1080/10408399609527761>
28. Osborne, N., Koplin, J.J., Martin, P.E., Gurrin, L.C., Lowe, A.J., Matheson, M.C., et al. (2011). Prevalence of challenge-proven IgE-mediated food allergy using population-based sampling and predetermined challenge criteria in infants. *Journal of Allergy and Clinical Immunology*, 127(1), 668–676.e2. <https://doi.org/10.1016/j.jaci.2011.01.039>
29. Rona, R.J., Keil, T., Summers, C., Gislason, D., Zuidmeer, L., Sodergren, E., et al. (2007). The prevalence of food allergy: a meta-analysis. *Journal of Allergy and Clinical Immunology*, 120(3), 638–646. <https://doi.org/10.1016/j.jaci.2007.05.026>
30. Sicherer, S.H., Sampson, H.A. (2010). Food allergy. *Journal of Allergy and Clinical Immunology*, 125(2, Suppl. 2), S116–S125. <https://doi.org/10.1016/j.jaci.2009.08.028>
31. Taylor, S. L., Lehrer, S. B. (1996). Principles and characteristics of food allergens. *Critical Reviews in Food Science and Nutrition*, 36 (SUPPL.), S91–S118. <https://doi.org/10.1080/10408399609527761>
32. Revyakina, V.A., Netrebenko, O.K. (2005). Allergic diseases in children and environment. Moscow: NewInform. — 240 p. (in Russian)
33. North, M.L., Ellis, A.K. (2011). The role of epigenetics in the developmental origins of allergic disease. *Annals of Allergy, Asthma & Immunology*, 106(5), 355–361. <https://doi.org/10.1016/j.anai.2011.02.008>
34. Strachan, D. P. (2000). Family size, infection and atopy: the first decade of the “hygiene hypothesis”. *Thorax*, 55(90001) S2–10. https://doi.org/10.1136/thorax.55.suppl_1.s2
35. Taylor, S. L., Hefle, S.L. (2001). Ingredient and labeling issues associated with allergenic foods. *Allergy*, 56(S67), 64–69. <https://doi.org/10.1034/j.1398-9995.2001.00920.x>
36. Taylor, S. L., S. L. Hefle. (2005). Food allergies and intolerances. In: *Modern Nutrition in Health and Disease*. Philadelphia: Williams and Wilkins. pp. 1512–1530.
37. Andjelkovic, U. (2020). Food Allergy & Food Allergens. *Reference Module in Food Science*. <https://doi.org/10.1016/b978-0-08-100596-5.22844-8>
38. Strid, J., Hourihane, J., Kimber, I., Callard, R., Strobel, S. (2004). Disruption of the stratum corneum allows potent epicutaneous immunization with protein antigens resulting in a dominant systemic Th2 response. *European Journal of Immunology*, 34(8), 2100–2109. <https://doi.org/10.1002/eji.200425196>
39. Howell, M.D., Kim, B.E., Gao, P., Grant, A.V., Boguniewicz, M., DeBenedetto, A., Schneider, L., Beck, L.A., Barnes, K.C., Leung, D.Y.M. (2007). Cytokine modulation of atopic dermatitis flagrin skin expression. *Journal of Allergy and Clinical Immunology*, 120(1), 150–155. <https://doi.org/10.1016/j.jaci.2007.04.031>
40. Perry, T.T., Conover Walker, M.K., Pomés, A., Chapman, M.D., Wood, R.A. (2004). Distribution of peanut allergen in the environment. *Journal of Allergy and Clinical Immunology*, 113(5), 973–976. <https://doi.org/10.1016/j.jaci.2004.02.035>
41. du Toit, G., Katz, Y., Sasieni, P., Mesher, D., Maleki, S.J., Fisher, H.R., Fox, A.T., Turcanu, V., Amir, T., Zadik-Mnuchin, G., Cohen, A., Livne, I., Lack, G. (2008). Early consumption of peanuts in infancy is associated with a low prevalence of peanut allergy. *Journal of Allergy and Clinical Immunology*, 122(5), 984–991. <https://doi.org/10.1016/j.jaci.2008.08.039>
42. Cummings, A.J., Knibb, R.C., King, R.M., Lucas, J.S. (2010). The psychosocial impact of food allergy and food hypersensitivity in children, adolescents and their families: a review. *Allergy*, 65(8), 933–945. <https://doi.org/10.1111/j.1398-9995.2010.02342.x>
43. Hefle, S.L. (1999). Impact of processing on food allergens. *Advances in Experimental Medicine and Biology*, 459, 107–119. https://doi.org/10.1007/978-1-4615-4853-9_8
44. Maleki, S.J., Chung, S.-Y., Champagne, E., T., Raufman, J.-P. (2000). The effects of roasting on the allergenic properties of peanut proteins. *Journal of Allergy and Clinical Immunology*, 106(4), 763–768. <https://doi.org/10.1067/mai.2000.109620>
45. Skolnick, H.S., Conover Walker, M.K., Barnes-Koerner, C., Sampson, H.A., Burks, W., Wood, R.A. (2001). The natural history of peanut allergy. *Journal of Allergy and Clinical Immunology*, 107(2), 367–374. <https://doi.org/10.1067/mai.2001.112129>
46. Wood, R.A. (2003). The natural history of food allergy. *Pediatrics*, 111(6 III), 1631–1637.
47. Lack, G. (2008). Epidemiologic risks for food allergy. *Journal of Allergy and Clinical Immunology*, 121(6), 1331–1336. <https://doi.org/10.1016/j.jaci.2008.04.032>
48. Yunginger, J.W., Nelson, D.R., Squillace, D.L., Jones, R.T., Holley, K.E., Hyma, B.A., Biedrzycki, L., Sweeney, K.G., Sturmer, W.Q., Schwartz, L.B. (1991). Laboratory investigation of deaths due to anaphylaxis. *Journal of Forensic Sciences*, 36(3), 857–865. <https://doi.org/10.1520/jfs13095j>
49. Keet, C.A., Wood, R.A. (2007). Food allergy and anaphylaxis. *Immunology and Allergy Clinics of North America*, 27(2), 193–212. <https://doi.org/10.1016/j.iac.2007.03.005>
50. Nica-Badea, D., Udristioiu, A., Andritioiu, C.V. (2018). Allergenicity risk assessment of foods derived from genetically modified

- fied crop plants. *Revue Francaise d'Allergologie*, 58(1), 29–34. <https://doi.org/10.1016/j.reval.2017.05.006> (In French)
51. Mills, E. N. C., E. Valovirta, C. Madsen, S. L. Taylor, S. Vieths, E. Ankliam, S. Baumgartner, P. Koch, R. W. R. Crevel, and L. Frewer. (2004). Information provision for allergic consumers – where are we going with food allergen labeling? *Allergy: European Journal of Allergy and Clinical Immunology*, 59(12), 1262–1268. <https://doi.org/10.1111/j.1398-9995.2004.00720.x>
52. FSA. Technical guidance: food allergen labeling information (EU Regulation 1169/2011) [Electronic resource: https://www.food.gov.uk/sites/default/files/media/document/fsa-food-allergen-labelling-and-information-requirements-technical-guidance_0.pdf Access date 15.04.2020]
53. FSIS Compliance Guidelines Allergens and Ingredients of Public Health Concern: Identification, Prevention and Control, and Declaration through Labeling, november 2015. [Electronic resource: <https://www.fsis.usda.gov/wps/wcm/connect/f9cb-b0e9-6b4d-4132-ae27-53e0b52e840e/Allergens-Ingredients.pdf?MOD=AJPERES> Access date 30.03.2020]
54. Technical regulations of the Customs Union TR CU022/2011 “Food products in terms of its labeling” (approved by the decision of the customs Union Commission of December 9, 2011 № 880). Moscow, – 2011. (in Russian)
55. Yurchak, Z.A., Kuznetsova, O.A., Starchikova, D. (2015). Prevention and minimization of a cross contamination of production food allergens. *Vsyo o myase*, 5, 19–21. (in Russian)
56. Crotty, M. P., Taylor, S.L. (2010). Risks associated with foods having advisory milk labeling. *Journal of Allergy and Clinical Immunology*, 125(4), 935–937. <https://doi.org/10.1016/j.jaci.2009.12.003>
57. Sikora, E., Górna, J. (2017). The Practical Aspects of Allergen Management in Meat Manufacturing in the United Kingdom. *Studia oeconomica posnaniensia*, 5(7), 162–176. <https://doi.org/10.18559/SOEP.2017.7.12>
58. Gordon, A., Williams, R. (2020). The role and importance of packaging and labeling in assuring food safety, quality and regulatory compliance of export products II: Packaging & labeling considerations. Chapter 7 in the book: Food Safety and Quality Systems in Developing Countries, 285–341. <https://doi.org/10.1016/b978-0-12-814272-1.00007-3>
59. Taylor, S.L., Hefle, S.L., Farnum, K., Rizk, S.W., Yeung, J., Barnett, M.E., Busta, F., Davis, S., Newsome, R., Shank, F.R., Bryant, C.M. (2007). Survey and evaluation of pre-FALCPA labeling practices used by food manufacturers to address allergen concerns. *Comprehensive Reviews in Food Science and Food Safety*, 6(2), 36–46. <https://doi.org/10.1111/j.1541-4337.2007.00016.x>
60. Voronovskaia, M.V. (2017). Food safety: food allergen management for processing enterprises. *Quality. Innovation. Education*, 11(150), 53–58. (in Russian)
61. Jackson, L.S., Al-Tajer, F.M., Moorman, M., DeVries, J.W., Tippet, R., Swanson, K.M.J., Fu, T.-J., Salter, R., Dunaif, G., Estes, S., Albillos, S., Gendel, S.M. (2008). Cleaning and Other Control and Validation Strategies to Prevent Allergen Cross-Contact in Food-Processing Operations. *Journal of Food Protection*, 71(2), 445–458. <https://doi.org/10.4315/0362-028X-71.2.445>
62. Kryuchenko, E. V. Chernukha, I.M., (2019). Some theoretical and practical aspects of the development of an allergen management system in the meat processing industry enterprises. *Collection of scientific papers of the international scientific and practical conference “Food Ingredients of Russia 2019”*. St. Petersburg: VNIIPD, 52–57. ISBN 978-5-901768-57-0 (in Russian)

AUTHOR INFORMATION

Elizaveta V. Kryuchenko — senior technician, Department of technical regulation and food safety systems, V. M. Gorbатов Federal Research Center for Food Systems of Russian Academy of Sciences. 109316, Moscow, Talalikhina str., 26. Tel.: +7-495-676-35-29, E-mail: l.kryuchenko@fncps.ru

ORCID: <https://orcid.org/0000-0002-5805-3055>

Yulya A. Kuzlyakina — candidate of technical sciences, chief researcher, Department of technical regulation and food safety systems, V. M. Gorbатов Federal Research Center for Food Systems of Russian Academy of Sciences. 109316, Moscow, Talalikhina str., 26. Tel.: +7-495-676-35-29, E-mail: yu.kuzlyakina@fncps.ru

ORCID: <https://orcid.org/0000-0002-2152-620X>

* corresponding author

Valentina S. Zamula — candidate of technical sciences, lead engineer, Department of technical regulation and food safety systems, V. M. Gorbатов Federal Research Center for Food Systems of Russian Academy of Sciences. 109316, Moscow, Talalikhina str., 26. Tel.: +7-495-676-35-29, v.zamula@fncps.ru

ORCID: <https://orcid.org/0000-0003-1634-1486>

Irina M. Chernukha — doctor of technical sciences, professor, Academician of Russian Academy of Sciences, Chief research scientist, Experimental clinic-laboratory «Biologically active substances of an animal origin», V. M. Gorbатов Federal Research Center for Food Systems of Russian Academy of Sciences. 109316, Moscow, Talalikhina str., 26.

Tel: +7-495-676-63-21, E-mail: imcher@inbox.ru

ORCID: <https://orcid.org/0000-0003-4298-0927>

All authors bear responsibility for the work and presented data.

All authors made an equal contribution to the work.

The authors were equally involved in writing the manuscript and bear the equal responsibility for plagiarism.

The authors declare no conflict of interest.

Received 14.05.2020 Accepted in revised 02.09.2020 Accepted for publication 25.09.2020

A SYSTEM APPROACH TO SIMULATION OF INDIVIDUAL FOOD PRODUCTS

Andrey B. Lisitsyn, Irina M. Chernukha, Marina A. Nikitina*

V. M. Gorbatov Federal Research Center for Food Systems of Russian Academy of Sciences, Moscow, Russia

Keywords: *simulation of living systems, individual food product, system approach, protein cleavage*

Abstract

There is no doubt that the further development in the field of nutrition is linked with personalization. Nutrition management with account for metabolism plays a key role in health strengthening and prevention of human diseases. The paper gives a review of studies associated with personalized nutrition. Personalized nutrition is inextricably linked with personalized food products. At present, however, mass production of personalized food products for individuals or small groups of people is unfeasible. The development of personalized food products requires both time and labor input, as well as multidisciplinary and profound knowledge in a wide spectrum of areas associated with biology, medicine, nutrition and food systems. Among the most important characteristics of modern science is the study of complex and super-complex organized objects such as the food system. These objects were studied previously but by the way of significant simplification of their structure. Investigation of objects with all variety and complexity of their organization requires not only new scientific ideas but also a new conceptual framework, new research methodology, new approaches to simulation of both products and physiological processes. In this study, the authors made an attempt to bring the theoretical view on an individual product closer to the complex task solution using the method of mathematical physiology. The intuitive conceptual model for a process of food design is shown with regard to the "health passport" of an individual, disease risk and gastrointestinal (GI) tract status. The differential equations of the concentration dynamics of protein, denatured protein and peptides in the human stomach are presented. The differential equations that describe the process of protein assimilation in the human stomach were solved in the simulation environment Simplex 3. The presented fragments of model realization show the possibility of virtual study on an effect of different indicators of the food nutritional value on the rate of digestion and the process of cleavage of complex components (proteins, fats and carbohydrates) to mono-structural elements depending on different state and influence factors.

Funding: The article was published as part of the research topic No. 0585–2019–0008 of the state assignment of the V. M. Gorbatov Federal Research Center for Food Systems of RAS.

Introduction

The idea of personalized medicine was presented for the first time by American biochemist Roger John Williams in the book "Biochemical Individuality: The Basis for the Genetotrophic Concept" [1] in 1956. In this book, R. J. Williams stressed the uniqueness of each person in terms of metabolism and requirements in nutritional microelements. Due to the "biochemical individuality", persons have their own nutrition requirements.

However, this idea was implemented only after human genome mapping in 2003 [2].

Nutrition management in chronic diet-related diseases can be improved based on data about the genome of a particular individual. To establish an interaction, the four-year Food4Me project (<http://www.food4me.org/>) financed by the European Commission was launched with the involvement of experts in the field of nutrition from different countries. The project envisaged individual recommendations on nutrition at three levels: 1) only a human diet; 2) a diet combined with the knowledge of human phenotype (measurable characteristics, such as height, weight, cholesterol level and so on); 3) a diet, phenotype and genotype (detection of the hereditary genetic identity of a person, for example, a gene variant linked with the weight gain). The European-wide

study (more than 1500 adults from Germany, Greece, Ireland, the Netherlands, Poland, Spain and UK) showed that personalized recommendations on nutrition were more effective for improving dietary behavior than traditional recommendations for population.

The results were presented at the project's final conference "Personalised nutrition: paving a way to better public health?" in Brussels on February 26, 2015. The study demonstrates that remote collection of phenotypic data and data on food consumption through the Internet is possible and allows obtaining consistent and reliable data [3,4,5].

It is worth emphasizing that personalized nutrition is inextricably linked with personalized food products. At present, mass production of personalized food products for individuals or small groups of people is unfeasible. Therefore, the development of personalized food products requires both time and labor input, as well as multidisciplinary and profound knowledge in a wide spectrum of areas associated with food products.

The fundamental trends that shape the world are analyzed and discussed in [6,7]. One of the megatrends is the availability of right personalized food products to prevent, mitigate diet-related diseases especially for vulnerable population groups including the elderly. At the same time, King et al. [7]

FOR CITATION:

Lisitsyn A. B., Chernukha I. M., Nikitina M. A. A system approach to simulation of individual food products. *Theory and practice of meat processing*. 2020;5(3): 12–17. <https://doi.org/10.21323/2414-438X-2020-5-3-12-17>.

believe that introduction of different types of microbiomes to cure people of certain diseases (personalized approach) will lead to food safety problems. Systematic research including the use of nutrigenomics, metabolomics and toxicogenomics is necessary to ensure safety of personalized products.

A process of designing personalized nutrition along the food chain is presented in [8] covering the following topics: 1) food raw materials and components; 2) food industry and gastronomy aspects; 3) digestion and microbiota; 4) food perception (Figure 1).

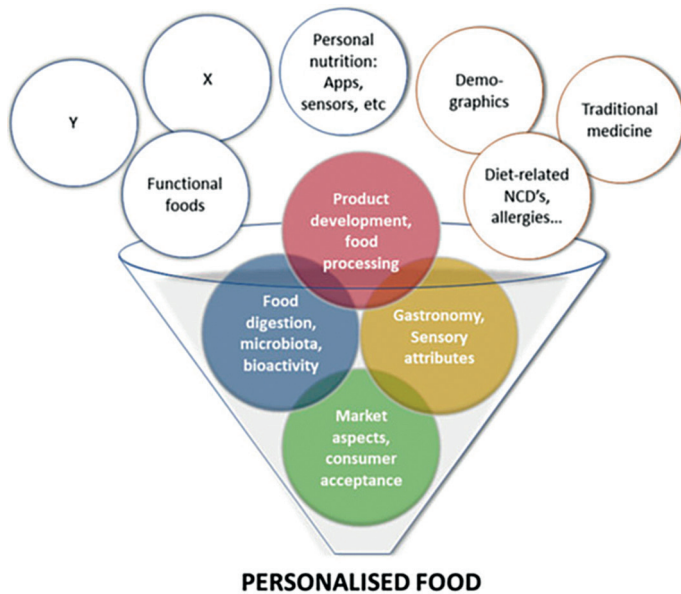


Figure 1. Individual approach to food [8]

The Biomolecula portal (<https://biomolecula.ru>) has the project “Science for life extension”, which points out that developed countries have faced the consequences of irrational nutrition such as obesity and type 2 diabetes mellitus over the last years [9] (Figure 2). It was established that longevity and the development of “deadly three” — cardiovascular, neurodegenerative and oncological diseases — depend on a human diet [10].

Medical data show the interrelation between nutrition and the most prevalent non-communicable diseases. In 2019, the research results of the scientific team from the USA, Norway and Spain were published [11]. The scientists analyzed data on the health state of the population from 195 countries and territories for the period from 1990 to 2017. They identified 92 diagnoses that finally led to irreversible deterioration of physical or mental abilities of patients. Among them were cardiovascular diseases (13), chronic respiratory diseases (6), cancers (35), diabetes, chronic kidney diseases, injuries and neurodegenerative diseases as well as vision and hearing impairments.

Analysis showed that age-related diseases accounted for more than half of health problems in all adult population of the world; however, data differed between countries and senility began at a very different age (Figure 3).

According to the data of 2017, the lowest SDI was in the Republic of the Niger (0.19), Somali (0.23), Afghanistan (0.29). SDI was 0.79 in Russia, 0.77 in Belarus, 0.7 in

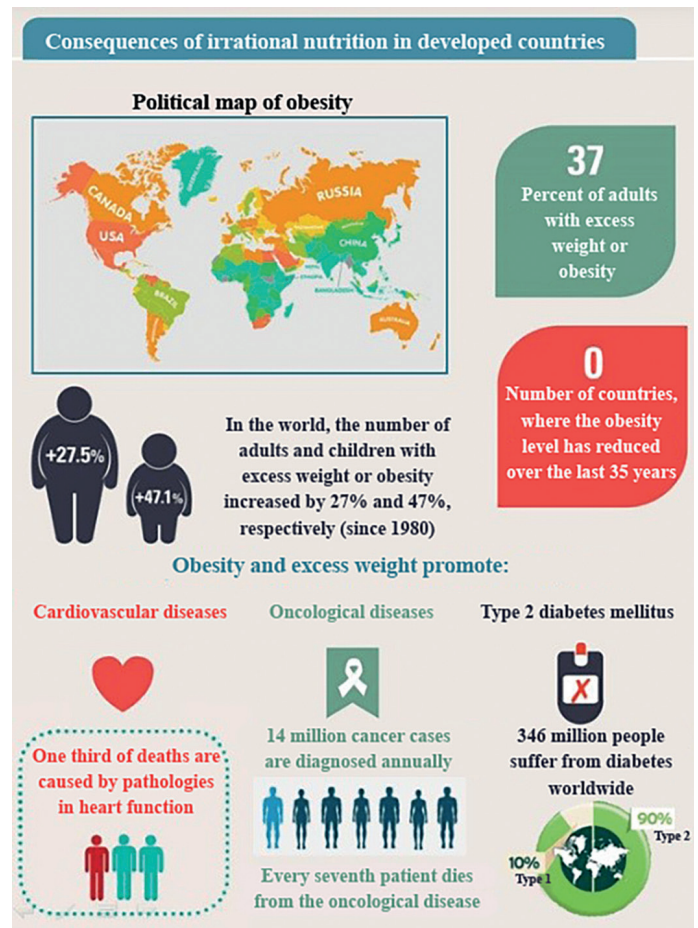


Figure 2. Consequences of unbalanced nutrition in developed countries: prevalence of obesity and type 2 diabetes mellitus. The excess weight and “western diet” contribute to the development of cardiovascular and oncological diseases, type 2 diabetes mellitus [biomolecula.ru]

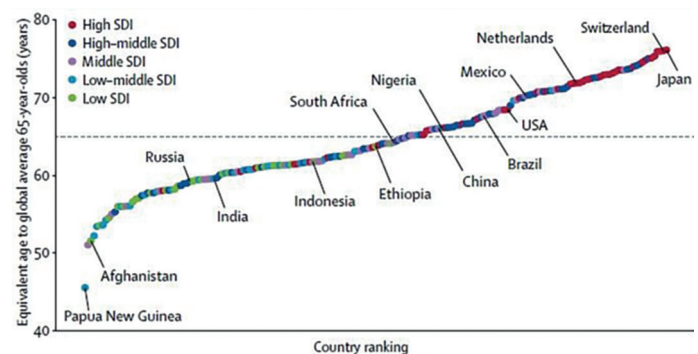


Figure 3: Comparison of the equivalent ages with global average index (65 years) across countries in 2017 [11]. SDI (Socio-demographic Index) — a summary measure of a degree of a country or region development expressed in values from 0 to 1 [12]

Armenia, 0.7 in Azerbaijan, 0.69 in Georgia, 0.74 in Kazakhstan, 0.6 in Kirgizstan, 0.62 in Uzbekistan. The highest SDI was in Denmark, Luxemburg (0.92), the Netherlands, Norway (0.91).

It is noted that although in developing countries (with low socio-demographic Index — SDI) longevity is considerably lower than in the developed ones, people still experience the burden of senility and accumulate age-related diseases. It just happens significantly earlier.

Chang A. et al. from the University of Washington Seattle have noticed that these incomparable results indicate that

an increase in longevity can be regarded in the old age both as additional opportunities and as a threat to well-being of a society in general depending on the age when the real age-related health problems arose in individuals and not on their actual age.

According to Ludwig von Bertalanffy [13], the system approach is an approach, in which any system (object) is regarded as an assembly of interrelated elements (components) having the output (target), input (resources), communication with the external environment and feedback. This is the most complex approach.

Among the most important peculiarities of modern science is the study (examination) of complex and super-complex organized objects. These objects were studied previously but by the way of significant simplification of their structure. Investigation of objects with all variety and complexity of their organization requires not only new scientific ideas but also a new conceptual framework, new research methodology [14].

The main advantage of simulation modeling compared to analytical is a possibility to solve more complex tasks. Simulation models allow quite easy consideration of factors such as the presence of discrete and continuous elements, non-linear characteristics of system elements, multiple random impacts and others, which often create difficulties in analytical investigations. At present, simulation modeling is the most effective method for investigation of complex systems and often is the only feasible method for obtaining information about system behavior especially at the stage of its design [14,15,16,17,18].

It should be mentioned that today an opportunity has arisen for the development and wider use of mathematical and information technologies and models in description of biological processes in particular linked with food production and consumption.

The authors of this paper propose using the system approach in simulation modeling of the development of individual food products with regard to the human "health passport", disease risk and gastrointestinal tract status.

The aim of the work is to use the system approach in the development of individual food products with detailing nutrient digestion in the human body.

Methodology of the research

The simulation modeling method enables solving tasks of complex system analysis including the tasks of assessing system structure variants, effectiveness of different algorithms of system management, effects of changes in different system parameters. Simulation modeling can also be used as the basis for structural, algorithmic and parametric synthesis of complex systems when it is necessary to create a system with given characteristics upon certain restrictions, which is optimal by several criteria of effectiveness assessment [14].

Simulation modeling was carried out in the program environment Simplex3 (<http://www.simplex3.net/>) [19,20].

Results

After human health screening and obtaining their "health passport", it is necessary to develop food products having regard to nutrient digestibility. To this end, it is necessary to formalize (translate to the mathematical language) processes occurring in the gastrointestinal tract using methods for modeling of living systems, including mathematical physiology methods [21,22].

The human gastrointestinal tract can be considered a natural biochemical reactor that ensures mechanical, thermal, enzymatic and microbiological processing of nutrients by their main components (proteins, fats, carbohydrates). The digestive process, therefore, consists in hydrolysis of nutrients (substrates) under the action of enzymes along the gastrointestinal tract and can be described by the system of Michaelis-Menten substrate-enzymatic reactions [23]. At the same time, it is necessary to take into account peculiarities of the human GI tract, first of all, pH, the presence of pathologies (for example, ulcer disease), food component composition, consistency and temperature.

We presented the structural-parametric model of the GI tract and structural -parametric model of a product separately in [24,25,26]. In the system approach, they will be examined as components of a single model.

A model of an individual food product requires not only consideration for food design according to indices of the nutritional and biological value but also the understanding of what part of nutrients in a food product will be assimilated by certain individuals with regard to their physiological characteristics. For this purpose, mathematical models of biological processes and equations of mathematical physics are used as a rule.

We will show the possibilities of simulation modeling by the example of protein cleavage and assimilation. To this end, at the beginning, we will describe multiple subsystems, organs, variables and links between them in the form of graphic presentation — the IDEF0 scheme (Figure 4).

Changes in the variables of the state of the certain GI tract segments can be described in finite differences by a sum of influencing components in the system of algebraic equations:

$$\Delta X_i = \sum_j^N w_{ij} \cdot f_{ij} \cdot \Delta X_j; i=1.20 \quad (1)$$

where f_{ij} is the influence function of the j^{th} influencing factor (enzymes, chemical substances of the influencing environment, mechanisms of substance transport) on the i^{th} parameter of the GI tract state; w_{ij} is the sign function of the direction of the influence of the j^{th} factor on the i^{th} , stimulating (+) or retarding (–).

Dynamics of the biochemical reactions of the enzymatic type as well as the biochemical and biomechanical processes of substance transport along the GI tract are described in [27] by the system of ordinary differential equations:

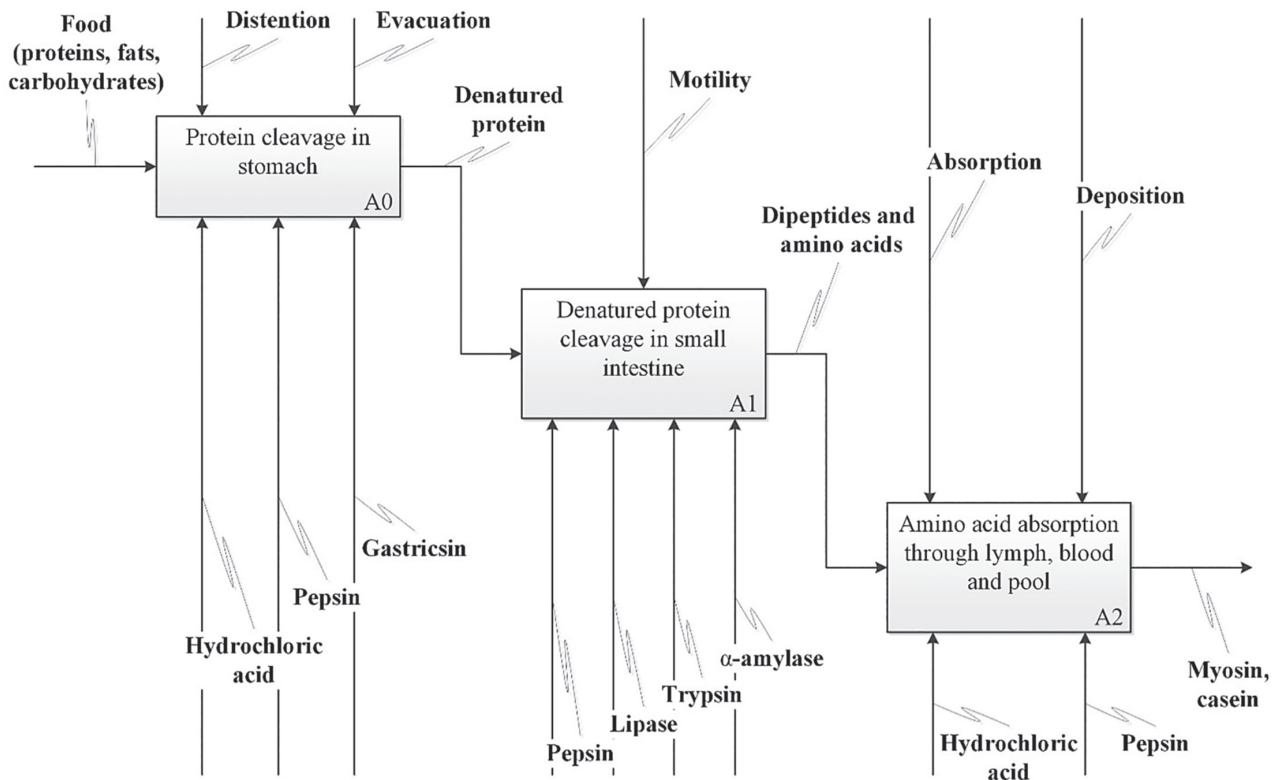


Figure 4. The scheme of protein cleavage; a blood pool — an organ or tissue with an ability to accumulate in its vessels a significant amount of blood, which can be used by the body when necessary (for example, liver, spleen, skin blood vessels and so on)

$$\frac{dX_i}{dt} = r \sum_{l=1}^{n_i} \sum_{j=1}^{23} \frac{w_{ij} \alpha_{jl} X_l Y_j}{(1 + \beta_{jl} X_l)}; i = \overline{1, 20} \quad (2)$$

where X , Y are variables of the system state and influencing factors of the biochemical environment, respectively; α_{jl} are coefficients of the intensity of the j^{th} influence of enzymes and transport mechanisms on the i^{th} parameter of the state; β_{jl} are coefficients of chemical reactions or substance transport; r are scale coefficients of rates of changes in variables in different GI tract segments.

Table 1 presents the used variables of the digestive system state and table 2 shows the factors of the biochemical environment of the GI tract and transport mechanisms.

Table 1. An example of description of food components transformed in the digestive process

Symbols	Product components determining digestive system state	Symbols	Product components determining digestive system state
<i>In stomach</i>		<i>In small intestine</i>	
x_1	Proteins	x_6	Denatured proteins
x_2	Fats (triglycerides)	x_7	Peptides
x_3	Carbohydrates (polysaccharides)	x_8	Dipeptides and amino acids
x_4	Denatured proteins	x_9	Fats (triglycerides)
x_5	Peptides	x_{10}	Emulsified fats
<i>In blood, lymph and pool</i>		x_{11}	
x_{17}	Amino acids	x_{12}	Fatty acids
x_{18}	Fatty acids	x_{13}	Carbohydrates (polysaccharides)
x_{19}	Triglycerides	x_{14}	oligosaccharides
x_{20}	Glucose	x_{15}	Disaccharides
		x_{16}	Glucose

Table 2. An example of analyzed factors of the GI tract environment

Symbols	Factors of the GI tract biochemical environment
y_1	Hydrochloric acid
y_2	Pepsin(ogens)
y_3	Bicarbonates of Brunner's glands
y_4	Bicarbonates of duodenum
y_5	Bicarbonates of pancreatic juice
y_6	Bile secretion (outflow to duodenum)
y_7	Enterokinase
y_8	Trypsinogen
y_9	Proteolytic enzymes of gastric juice
y_{10}	Peptidase
y_{11}	Dipeptidase
y_{12}	Lipase
y_{13}	Monoglyceride lipase
y_{14}	α -Amylase
y_{15}	Oligosaccharidase
y_{16}	Dissaccharidase
y_{17}	Electrolytes in intestine
y_{18}	Ca^{++}
Transport mechanisms	
y_{19}	Gastric distention
y_{20}	Gastric evacuation
y_{21}	Intestinal motility
y_{22}	Absorption
y_{23}	Deposition

In relation to the symbols (Table 1 and Table 2) and IDEF0 scheme, equation (2):

- for dynamics of the concentration of entered proteins (x_1) in the stomach with regard to the factors of protein denaturation by hydrochloric acid (y_1), gastric distention (y_{19}) and evacuation (y_{20}) is described in the expanded form by the following differential equation:

$$\frac{dx_1}{dt} = x_1 - \frac{\alpha_{1.19}x_1}{1 + \beta_{1.19}x_1}y_{19} - \frac{\alpha_{1.1}x_1}{1 + \beta_{1.1}x_1}y_1 - \frac{\alpha_{1.20}x_1}{1 + \beta_{1.20}x_1}y_{20} \quad (3)$$

- for dynamics of the concentration of denatured protein in the stomach as a result of an impact of hydrochloric acid (y_1) on entered protein (x_1), reduction of the concentration of denatured protein (x_4) due to an impact of pepsin (y_2) and partial evacuation into small intestine (y_{20}) has the form:

$$\frac{dx_4}{dt} = \frac{\alpha_{1.1}x_1}{1 + \beta_{1.1}x_1}y_1 - \frac{\alpha_{2.4}x_4}{1 + \beta_{2.4}x_4}y_2 - \frac{\alpha_{20.4}x_4}{1 + \beta_{20.4}x_4}y_{20} \quad (4)$$

- for dynamics of the peptide (x_5) concentration in the stomach reflects their accumulation upon cleavage of denatured protein (x_4) by pepsin (y_2) and reduction as a result of the further evacuation (y_{20})

$$\frac{dx_5}{dt} = \frac{\alpha_{2.4}x_4}{1 + \beta_{2.4}x_4}y_2 - \frac{\alpha_{20.5}x_5}{1 + \beta_{20.5}x_5}y_{20} \quad (5)$$

The differential equations of the dynamics of fat and carbohydrate cleavage are formulated in a similar way.

The initial data of the task were the coefficients of enzyme activities and transport mechanisms $\alpha_1 = 0.03$, $\alpha_2 = 0.01$, $\alpha_{20} = 0.001$; the coefficients of the chemical reaction kinetics or substance transfer $\beta_1 = 1$, $\beta_2 = 1$, $\beta_{20} = 0.01$; the initial protein concentration was $x_1 = 0.1$ (Figure 5).

In the stomach, a decrease in the concentration of the protein substrate X1 is observed, which is transformed into the denatured form and/or evacuated from the stomach. At the same time, the concentration of denatured protein increases with the further cleavage to peptides X5.

The presented fragment of model realization in the universal simulation model Simplex 3 shows a possibility of virtual study on the influence of different food nutritional value indicators on the rate of digestion and the process of cleavage of complex components to mono-structural elements depending on different state and influence factors.

Conclusion

An advantage of personalized nutrition is indisputable. Nowadays, however, mass production of personalized food products for individuals or small groups of people is unfeasible. The development of personalized food products requires not so much time and labor input as multidisciplinary and profound knowledge in a wide spectrum of areas associated with nutrition, biochemistry, physiology and food technologies. It is necessary to use the system approach with an integrated and detailed examination of a complex food system. The authors made an attempt to bring the theoretical view on an individual product closer to a complex task solution using the method of mathematical physiology. The process of protein cleavage in the form of the IDEF0 scheme is presented. The possibility to simulate the process of protein assimilation in the human stomach is shown in the simulation environment Simplex 3.

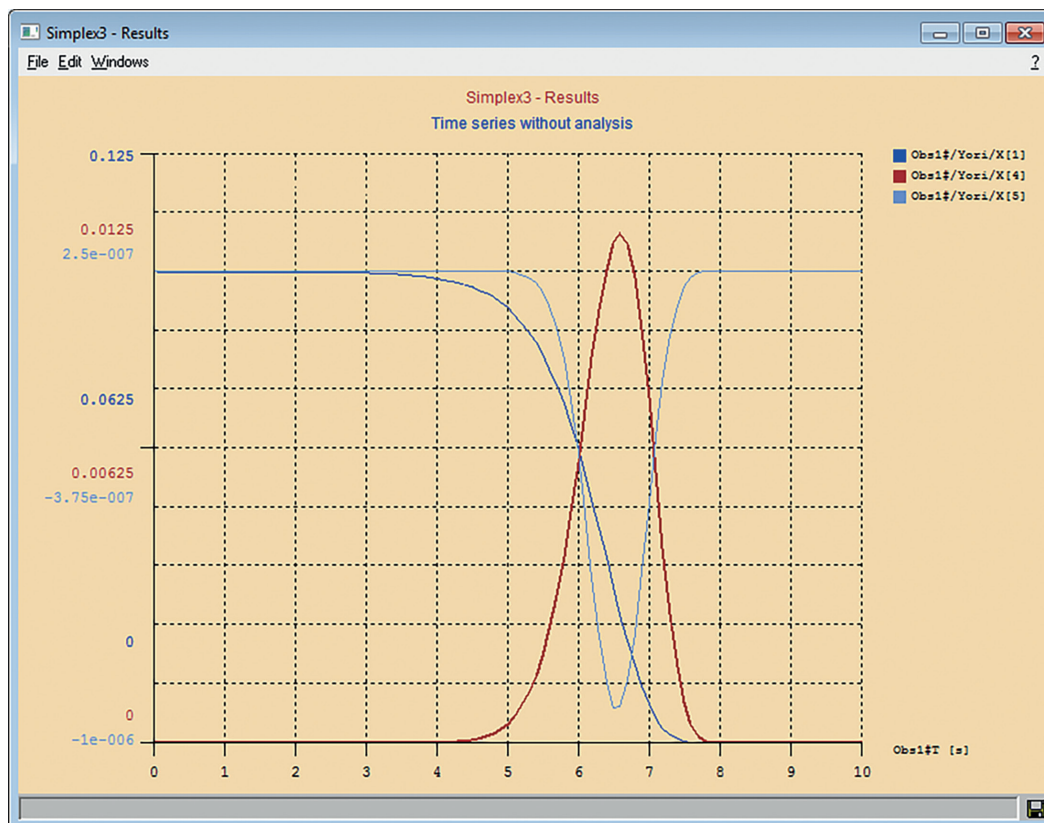


Figure 5. Changes in the concentration of proteins X1, denatured protein X4, and peptides X5 in the stomach during the digestive process

REFERENCES

- Williams, R.J. (1956). *Biochemical Individuality: The Basis for the Genetotropic Concept*. New York: John Wiley & Sons. — 272 p.
- Levy, S., Sutton, G., Ng, P.C., Feuk, L., Halpern, A.L., et al. (2007). The Diploid Genome Sequence of an Individual Human. *Plos Biology*, 5(10), e254. <https://doi.org/10.1371/journal.pbio.0050254>
- Celis-Morales, C., Mathers, J.C., Cibney, M., Walsh, M., et al. (2015). White paper on personalised nutrition — paving a way to better population health. Technical Report. — 101 p. <https://doi.org/10.13140/RG.2.2.13147.16166>
- EU-funded Food4Me project paves way for personalised nutrition to better public health. *Food Today*. — 2015. — Vol. 97.
- Celis-Morales, C., Livingstone, K.M., Marsaux, C.F.M., et al. (2015). Design and baseline characteristics of the Food4Me study: a web-based randomised controlled trial of personalised nutrition in seven European countries. *Genes&Nutrition*, 10(1), article number 450. <https://doi.org/10.1007/s12263-014-0450-2>
- Franklin, D., Andrews, J. (2012). *Megachange: the world in 2050*. Hoboken: John Wiley & Sons. — 320 p. ISBN 1118180445
- King, T., Cole, M., Farber, J.M., Eisenbrand, G., Zabaras, D., Fox, E.F., Hill, J.P. (2017). Food safety for food security: Relationship between global megatrends and developments in food safety. *Trends in Food Science & Technology*, 68, 160–175. <https://doi.org/10.1016/j.tifs.2017.08.014>
- Ueland, Ø., Altintzoglou, T., Kirkhus, B., Lindberg, D., Rognså, G.H., Rosnes, J.T., Rud, I., Varela, P. (2020). Perspectives on personalised food. *Trends in Food Science & Technology*, 102, 169–177. <https://doi.org/10.1016/j.tifs.2020.05.021>
- Neeha, V.S., Kinth, P. (2013). Nutrigenomics research: a review. *Journal Food Science and Technology*, 50(3), 415–428. <https://doi.org/10.1007/s13197-012-0775-z>
- Willett, W.C. (2002). Balancing life-style and genomics research for disease prevention. *Science*, 296(5568), 695–698. <https://doi.org/10.1126/science.1071055>
- Chang, A.Y., Skirbekk, V.F., Tyrovolas, S., Kassebaum, N.J., Dieleman, J.L. (2019). Measuring population ageing: an analysis of the Global Burden of Disease Study 2017. *The Lancet Public Health*, 4, 3, e159–e167. [https://doi.org/10.1016/S2468-2667\(19\)30019-2](https://doi.org/10.1016/S2468-2667(19)30019-2)
- Socio-demographic Index (SDI) [Electronic resource: <http://www.healthdata.org/taxonomy/glossary/socio-demographic-index-sdi>. Access date 14.08.2020] (in Russian)
- von Bertalanffy, L. (1950). The Theory of Open Systems in Physics and Biology. *Science*, 111(2872), 23–29. <https://doi.org/10.1126/science.111.2872.23>
- Sovetov, B. Ya., Yakovlev, S.A. (2017). Systems simulation. Moscow: Yuriat. — 343 p. (in Russian)
- Buslenko, N.P. (1988). *Complex systems simulation*. Moscow: Science. — 400 p. (in Russian)
- Vavilov, A.A. (1983). *Simulation modeling of production systems*. Moscow: Mashinostroenie. — 417 p. (in Russian)
- Kosturiak, J., Gregor, M. (1995). *Simulation von productions-systemen*. Berlin: Springer. — 180 p. ISBN 978-3-211-82701-7, <https://doi.org/10.1007/978-3-7091-9413-3> (In Germany)
- Shannon, R.E. (1975). *Systems simulation. The art and science*. New Jersey: Prentice-Hall. — 387 p.
- Schmidt, B. (2001). *The art of modelling and simulation: introduction to the simulation system Simplex3*. Ghent.: SCS Europa BVBA. — 500 p. ISBN 3-936150-26-5
- Ivashkin, Yu.A. (2016). *Multi-agent modeling in simulation system Simplex3*. Moscow: Knowledge Laboratory — 360 p. ISBN 978-5-906828-72-9 (in Russian)
- Keener, J., Sneyd, J. *Mathematical Physiology*. New York: Springer-Verlag. — 547 p. ISBN 978-1-4899-8670-2, <https://doi.org/10.1007/978-0-387-75847-3>
- Murray, J. D. (2002). *Mathematical Biology*. New York: Springer-Verlag. — 551 p. ISBN 978-0-387-95228-4
- Michaelis, L., Menten, M. L. (1913). Die Kinetik der Invertinwirkung. *BiochemischeZeitschrift*, 49, 333–369. (In Germany)
- Lisitsyn, A.B., Chernukha, I.M., Nikitina, M.A. (2019). Development of a personalized meat product using structural-parametric modeling. *Theory and practice of meat processing*, 4(3), 11–18. <https://doi.org/10.21323/2414-438X-2019-4-3-11-18>
- Ivashkin, Yu.A., Nikitina, M.A. (2018). The concept of biological compatibility in optimization of a human diet. *Science intensive technologies*, 19, 3, 33–45. (in Russian)
- Ivashkin, Yu.A., Nikitina, M.A. (2017). Multi-agent simulation model of human gastrointestinal tract. *Mathematical Methods in Technique and Technologies — MMTT*, 5, 77–84. (in Russian)
- Polenov, S.A., Troitskaya, V.K., Vershinina, E.A. (2003). Regulation of the digestive process: the main mechanisms and their computer simulation. *Russian Journal of Gastroenterology, Hepatology, Coloproctology, (Application)* 4, 25–29. (in Russian)

AUTHOR INFORMATION

Andrey B. Lisitsyn — doctor of technical sciences, professor, Academician of the Russian Academy of Sciences, Scientific supervisor, V. M. Gorbato Federal Research Center for Food Systems of Russian Academy of Sciences. 109316, Moscow, Talalikhina str., 26. Tel: +7-495-676-95-11, E-mail: info@fncps.ru
ORCID: <https://orcid.org/0000-0002-4079-6950>

Irina M. Chernukha — doctor of technical sciences, professor, Academician of the Russian Academy of Sciences, leading research scientist of Experimental clinic — laboratory «Biologically active substances of an animal origin, V. M. Gorbato Federal Research Center for Food Systems of Russian Academy of Sciences. 109316, Moscow, Talalikhina str., 26. Tel: +7-495-676-97-18, E-mail: imcher@inbox.ru
ORCID: <https://orcid.org/0000-0003-4298-0927>

Marina A. Nikitina — candidate of technical sciences, docent, leading scientific worker, the Head of the Direction of Information Technologies of the Center of Economic and Analytical Research and Information Technologies, V. M. Gorbato Federal Research Center for Food Systems of Russian Academy of Sciences. 109316, Moscow, Talalikhina str., 26 Tel: +7-495-676-92-14, E-mail: m.nikitina@fncps.ru
ORCID: <https://orcid.org/0000-0002-8313-4105>

* corresponding author

All authors bear responsibility for the work and presented data.

All authors made an equal contribution to the work.

The authors were equally involved in writing the manuscript and bear the equal responsibility for plagiarism.

The authors declare no conflict of interest.

Received 09.08.2020 Accepted in revised 15.09.2020 Accepted for publication 25. 09.2020

STUDY OF THE FUNCTIONAL PRODUCT'S PROTEIN COMPOUNDS DIGESTION FEATURES

Ekaterina R. Vasilevskaya*, Anastasiya G. Akhremko, Ekaterina K. Polishchuk, Liliya V. Fedulova

V. M. Gorbatov Federal Research Center for Food Systems of Russian Academy of Sciences, Moscow, Russia

Keywords: digestion, biocorrecting meat product, proteins, peptides, one-dimensional electrophoresis

Abstract

The aim of the study was to investigate the transformation of meat product's proteins from pig hearts and aortas during enzymatic hydrolysis in an *in vitro* model of the gastrointestinal tract. The model consisted of three phases simulating digestion processes: "oral cavity" phase (α -amylase, pH 7.0; 2 min), "stomach" phase (pork pepsin, pH 3.0; 120 min), "intestine" phase (pork pancreatin, pH 7.0; 130 min). The product was sequentially subjected to hydrolysis, at the end of each phase, samples were taken to determine the protein concentration (biuret method) and visualize the protein fractions (one-dimensional electrophoresis). A significant increase in protein concentration at the "stomach" phase was revealed by 3.2 times, and the absolute content by 4.6 times. At the "intestine" phase, a decrease in the number of peptide complexes with copper ions by 1.8 times, the absolute protein content by 8.5% was revealed. The noted tendency was confirmed by electrophoretic studies — at the stage, simulating digestion in the stomach, the products of meat product's proteins hydrolysis were visualized; at the "intestine" phase, a low expression of protein fractions in the range of more than 10 kDa is shown. The maximum hydrolysis of protein compounds at the "stomach" phase to poly- and oligopeptides was confirmed, continuing at the "intestine" stage with the accumulation of free amino acids. This methodology makes it possible to visualize the products of hydrolysis of proteins in a meat product at all stages of the model and to monitor changes in protein concentration in the system.

Funding: The research was supported by state assignment of V. M. Gorbatov Federal Research Centre for Food Systems of RAS, scientific research No. 013.20.05.

Introduction

Animal food products' proteins are of particular interest today due to the fact that such compounds are the main source of biologically active components.

Today bioactive components of foods are considered dietary elements that have a measurable biological effect (for example, antioxidant, immunomodulatory, antihypertensive, osteoprotective, hypolipemic, opiate, etc.) and have a beneficial effect on human health [1,2,3]. During different phases of industrial processing and digestion, bioactive peptides with a wide range of biological effects can be released from protein [4,5].

It is known that bioactive peptides are naturally formed in mammals in the gastrointestinal tract during the metabolism of dietary meat proteins. Consumed proteins derived from raw meat are attacked by digestive enzymes secreted by the stomach, such as pepsin, and, in the small intestine, trypsin, chymotrypsin, elastase, and carboxypeptidase undergo proteolysis. To study peptides of meat products that potentially have functional effects, a simulated digestive system of the gastrointestinal tract can be used to reproduce the processes of generating peptides similar to those released during physiological digestion. The process that simulates digestion of the gastrointestinal tract is based on enzymatic hydrolysis using various commercial exogenous proteinases obtained from animal tissues (pepsin and trypsin), plants (papain, ficin and bromelain) and microbial sources (alkalase, collagenase or proteinase, etc.) [6,7].

The number of studies that use the methodology of modeling gastrointestinal digestion *in vitro* to study bioactive peptides from food proteins are increasing [8,9,10,11,12,13]. Using these methods, researchers are trying to isolate bioactive peptides that can be produced in our body after eating a certain food or dietary protein, and determine its activity.

In this article, we will attempt to understand meat products' proteins transformation using the example of a biocorrecting product based on pigs' hearts and aortas after passing through the digestive tract.

Materials and methods

The object of the study was an experimental development — a meat product made on the basis of pig hearts and aortas in accordance with [14,15], containing functional proteins with biocorrecting properties [16,17,18].

The functional product is a sterilized canned minced meat, it contains $17.5 \pm 0.9\%$ protein, $3.8 \pm 0.1\%$ fat, $0.30 \pm 0.01\%$ sodium chloride and $2.3 \pm 0.2\%$ starch.

To simulate the digestion process of the experimental product, a static model of digestion was used in accordance with [19]. Pate crushed in a mortar to a particle size of 3 ± 0.5 mm in amount of 10 g was placed in I imitating liquid ("oral cavity", pH 7, volume 7 ml), consisting of 0.5 ml of α -amylase (30 U/mg, "PanReac AppliChem", Germany), 25 μ l of calcium chloride (0.3 M), 10.6 ml of phosphate-salt buffer ("Sigma-Aldrich", USA) and 0.09

ml of hydrochloric acid (6 M, “Sigma-Aldrich”, USA). The mixture was then thoroughly mixed for 2 minutes and a sample was taken for further studies. Next, 10 ml of the mixture was placed in II imitating liquid (“stomach”, pH 3, volume 15 ml), consisting of 1.6 ml of pig pepsin solution (250 U / mg, “PanReac AppliChem”, Germany), 5 ml of calcium chloride (0.3 M, “Sigma-Aldrich”, USA), 0.8 ml of hydrochloric acid (1 M) and 10.0 ml of phosphate-salt buffer. The mixture was stirred using a magnetic stirrer for 120 minutes, and samples were taken for further studies at the end of the incubation time. Then 20 ml of gastric hummus was added to the III imitating liquid (“intestines”, pH 7, volume 20 ml) containing 5.0 ml of porcine Pancreatin solution (50.5 U / mg, “PanReac AppliChem”, Germany), 1.5 ml of canned bovine bile (“Samson-med”, Russia) and 40 ml of 0.3 M calcium chloride, 10.6 ml of phosphate-salt buffer. To neutralize the mixture of a neutral pH value, 0.6 ml of 1 M sodium hydroxide (Sigma-Aldrich, USA) and 986 µl of distilled water were added. The mixing time was 130 minutes, then samples were also taken. In parallel, similar procedures were performed for all stages of the model without adding the test sample to obtain control samples.

All samples were taken in plastic tubes of the Eppendorf type and placed in a freezer at a temperature of minus $(40 \pm 1)^\circ\text{C}$ to inactivate the enzymatic activity for at least 60 minutes. After defrosting, the samples were centrifuged using a CM-6M centrifuge (ELMI, Latvia) at 3500 rpm for 5 minutes.

The protein concentration in the supernatant of the samples after centrifugation was measured by biuret method in accordance with [20] on Biochem SA photometer (HTI, USA).

For electrophoretic analysis, $50,00 \pm 0,05$ ml of the sample's supernatant was taken into a microcentrifuge tube, 50 µl of solubilizing solution was added (Glycerol 10%, β-mercaptoethanol 0.01 M, Bromphenol blue 0.02%, Tris-HCl 0.5 M, SDS2% — “Sigma-Aldrich”, USA) and kept in a thermostat at 95°C for 5 minutes. The resulting solution was centrifuged in an Eppendorf 5402r centrifuge at 14,000 rpm for 8 minutes. The supernatant was used for further analysis.

Analysis of the fractional composition was carried out by denaturing electrophoresis in 10% polyacrylamide gel in the presence of sodium dodecyl sulfate using a Helli-con “VE-10” electrophoretic chamber, at a constant current and voltage of 55 V and 130 V, for 2 hours. As a solution of standards, a marker consisting of standard preparations with a molecular weight is used 250, 150, 100, 70, 50, 40, 30, 20 kDa (“Thremo”, Latvia). Visualization was performed with Coomassie R250 solution and silver nitric acid (“Sigma-Aldrich”, USA).

Results and discussion

Proteins' biotransformation in the product was evaluated by quantifying the formation of protein peptide bonds

with divalent copper ions during modeling and visualization of protein compounds after gradual digestion by digestive enzymes.

According to the study results, it was found that during modeling stage I, which recreates the conditions of product's digestion in the “oral cavity”, there were no significant changes in the protein concentration (Figure 1). The main transformations were revealed after the second phase corresponding to digestion in the “stomach” — the protein concentration increased by 3.2 times, and the absolute protein content increased by 4.6 times. At stage III, after passing the simulated “intestine” fluid, the protein concentration in the system decreased by 1.8 times, but the absolute protein content decreased by only 8.5%, relative to stage II.

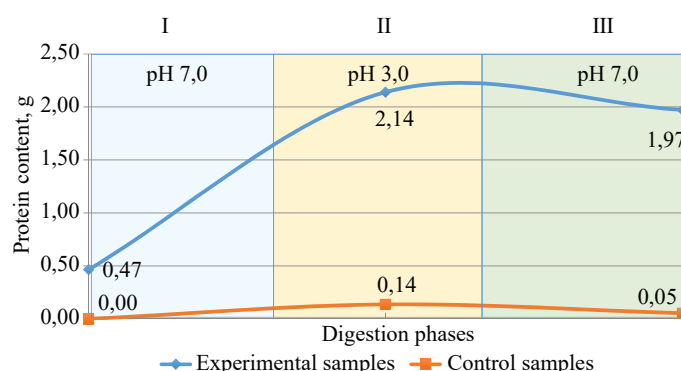


Figure 1. Protein content after digestion model phases

When one-dimensional electrophoresis was stained with Coomassie solution, the preservation of product's major fractions with molecular weights in range from 35 to 37 kDa and 13 kDa was shown (Figure 2A, track 1), and fractions corresponding to oral enzymes were also identified. When visualizing with silver nitrate, the presence of an almost complete spectrum of the initial product can be noted, with the addition of oral enzymes fractions (Figure 2B, track 1).

On the second track of the electrophoregram, the result of enzymatic hydrolysis of product proteins was demonstrated. Coomassie staining revealed a protein front in the range of molecular weights less than 12 kDa, which was not previously detected in the product, as well as minor fractions (42–48 kDa) corresponding to “stomach” enzymes (Figure 2A, track 2). Nitric silver as a dye allowed us to detect the presence of protein fractions in the range of molecular weights from 250 to 80 kDa, from 40 to 25 kDa, and from 17 to 14 kDa, which correspond to the proteins initially present in the product (Figure 2B, track 2).

Similar results were found when visualizing product's protein profile at the end of the third phase of digestion (“intestine”). After Coomassie staining, low-molecular compounds (less than 12 kDa) and enzymatic minor bands in the molecular weight range of 28–30 kDa were also visualized (Figure 2A, track 3). The dye with silver nitric acid also showed the presence of protein bands fixed in the sample obtained after the “intestine” stage (Figure 2B, tracks 2 and 3).

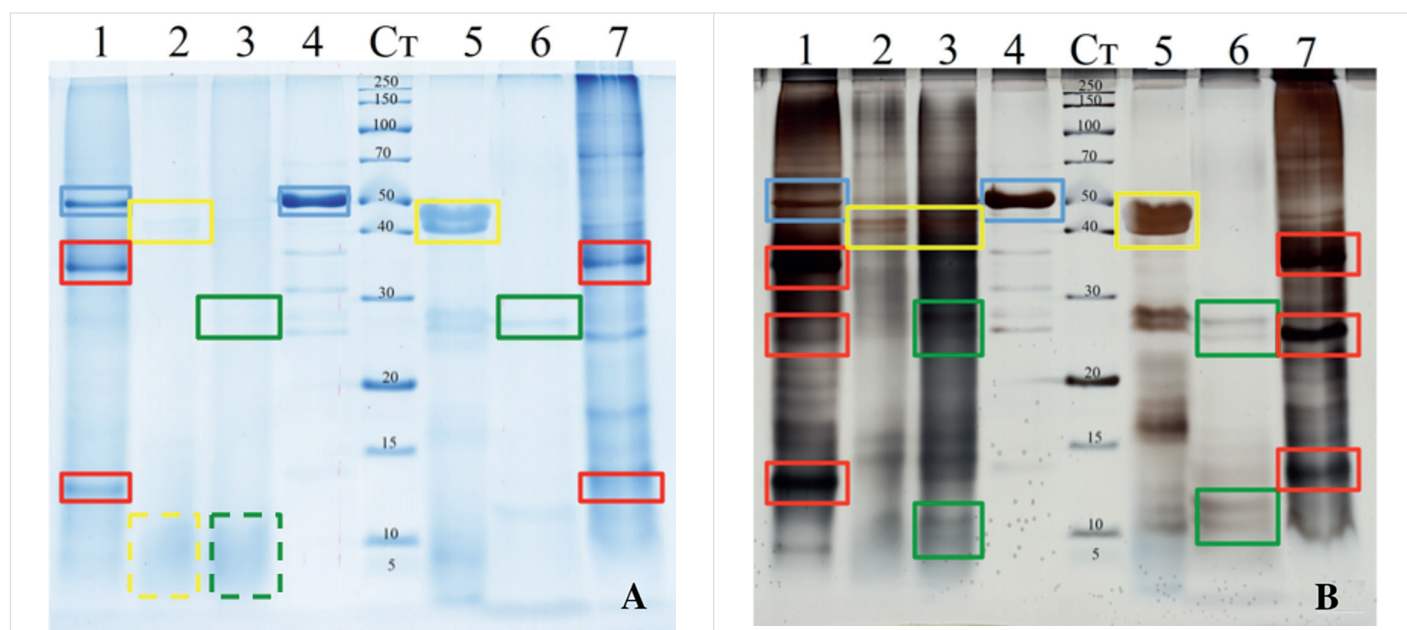


Figure 2. One-dimensional electrophoregrams of the studied samples. A — Coomassie color; B — silver nitric acid color.

Symbols:

- 1 — I phase — imitation of digestion in the “oral cavity”;
- 2 — II phase — imitation of digestion in the “stomach”;
- 3 — III phase — imitation of digestion in the “gut”;
- 4 — control sample for phase I;

- Ct — standard for molecular weights: 250, 150, 100, 70, 50, 40, 30, 20 kDa (top to bottom)
- 5 — control sample for phase II;
- 6 — control sample for phase III;
- 7 — product before the experiment.

The revealed tendency to decrease the concentration of protein and its absolute content is due to the fact that the “stomach” phase is active hydrolysis of product proteins by pig pepsin, which cleaves the central peptide bonds in protein molecules. At this phase, a large number of poly- and oligopeptides and a small amount of free amino acids are formed. The decrease in the number of peptide complexes with divalent copper ions at the “intestine” stage is due to the specificity of the action of trypsin and chymotrypsin, which destroy them with the formation of carboxyl groups of amino acids, as well as free amino acids [21,22].

As expected, the one-dimensional electrophoregram also visualized hydrolysis of meat product proteins at the stage simulating digestion in the stomach. When simulating digestion in the “intestine”, the marked decrease in the number of peptide complexes with copper ions (by 8.5%) correlates with a low expression of protein fractions in the

range of more than 10 kDa and confirms the further metabolism of protein compounds — the conversion of peptides to free amino acids.

Conclusion

The prospects of using a complex technique for studying the digestibility of functional products’ proteins, including a three-stage enzymatic model of digestion with sequential sampling to determine the protein concentration at each stage and visualization of individual components on one-dimensional electrophoresis, were proved. The demonstrated results of the study indicate the accumulation of a peptide pool during digestion, and special attention should be paid to “stomach” and “intestine” phases.

The next step in the study of bioactive peptides obtained in the meat product proteins’ proteolysis process, that can be identified, and its’ absorption study at the intestinal level in the *in vitro* system using cell cultures.

REFERENCES

1. Bechaux, J., Gatellier, P., Le Page, J.F., Drillet, Y., Sante-Lhoutellier, V. (2019). A comprehensive review of bioactive peptides obtained from animal byproducts and their applications. *Food and Function*, 10(10), 6244–6266. <https://doi.org/10.1039/c9fo01546a>
2. Albenzio, M., Santillo, A., Caroprese, M., Della Malva, A., Marino, R. (2017). Bioactive Peptides in Animal Food Products. *Foods*, 6(5), 35. <https://doi.org/10.3390/foods6050035>
3. Kupaeva, N.V., Kotenkova, E.A. (2019). Analysis of the antioxidant capacity of farm animal raw materials. *Vsyo o myase*, 5, 34–37. <https://doi.org/10.21323/2071-2499-2019-5-34-37> (in Russian)
4. Chernukha, I.M., Afanasyev, D.A., Mashentseva, N.G., Vostrikova, N.L. (2019). Biologically active peptides as a product of microbial fermentation of raw meat and finished meat products: review. Part 1. General information about biologically active peptides of meat and meat products. *Theory and practice of meat processing*, 4(4), 12–16. <https://doi.org/10.21323/2414-438X-2019-4-4-12-16> (in Russian)
5. Chernukha, I.M., Afanasyev, D.A., Mashentseva, N.G., Vostrikova, N.L. (2020). Biologically active peptides as a product of microbial fermentation of raw meat and finished meat products: review. Part 2. Functionality of bioactive meat peptides. *Theory and practice of meat processing*, 5(2), 12–19. <https://doi.org/10.21323/2414-438X-2020-5-2-12-19> (in Russian)
6. Fu, Y., Zhang, Y., Soladoye, O.P., Aluko, R.E. (2019). Maillard reaction products derived from food protein-derived peptides: insights into flavor and bioactivity. *Critical reviews in food science and nutrition*, 18, 1–14. <https://doi.org/10.1080/10408398.2019.1691500>
7. Lafarga, T., O'Connor, P., Hayes, M. (2015). In silico methods to identify meat-derived prolyl endopeptidase inhibitors. *Food*

Chemistry, 175, 337–343. <https://doi.org/10.1016/j.foodchem.2014.11.150>

8. Kong, F., Singh, R.P. (2010). A human gastric simulator (HGS) to study food digestion in human stomach. *Journal of Food Science*, 75(9), E627–E635. doi:10.1111/j.1750-3841.2010.01856.x

9. Picariello, G., Miralles, B., Mamone, G., Sánchez-Rivera, L., Recio, I., Addeo, F., Ferranti, P. (2015). Role of intestinal brush border peptidases in the simulated digestion of milk proteins. *Molecular Nutrition and Food Research*, 59(5), 948–956. <https://doi.org/10.1002/mnfr.201400856>

10. Vieira, E.F., das Neves, J., Vitorino, R., Dias da Silva, D., Carmo, H., Ferreira, I.M. (2016). Impact of in vitro Gastrointestinal Digestion and Transepithelial Transport on Antioxidant and ACE-Inhibitory Activities of Brewer's Spent Yeast Autolysate. *Journal of Agricultural and Food Chemistry*, 64(39), 7335–7341. <https://doi.org/10.1021/acs.jafc.6b02719>

11. Mora, L., Bolumar, T., Heres, A., Toldrá, F. (2017). Effect of cooking and simulated gastrointestinal digestion on the activity of generated bioactive peptides in aged beef meat. *Food and Function*, 8(12), 4347–4355. <https://doi.org/10.1039/C7FO01148B>

12. Aspri, M., Leni, G., Galaverna, G., Papademas, P. (2018). Bioactive properties of fermented donkey milk, before and after in vitro simulated gastrointestinal digestion. *Food Chemistry*, 268, 476–484. <https://doi.org/10.1016/j.foodchem.2018.06.119>

13. Chakrabarti, S., Guha, S., Majumder, K. (2018). Food-Derived Bioactive Peptides in Human Health: Challenges and Opportunities. *Nutrients*, 10(11), 1738. <https://doi.org/10.3390/nu10111738>

14. Lisitsyn, A.B., Chernukha, I.M., Fedulova, L.V., Kotenkova, E. A. Functional meat product and its manufacture method. Patent RF, no. 2550649, 2015. (in Russian)

15. Kotenkova E. A. (2015). Application of biotechnological and proteomic methods in the development of food products with

hypolipidemic and vasoprotective effects. Author's abstract of the dissertation for the scientific degree of Candidate of Technical Sciences. Moscow: VNIIMP. — 22 p. (in Russian)

16. Chernukha, I., Fedulova, L., Kotenkova, E., Akhremko, A. (2018). Hypolipidemic action of the meat product: in vivo study. *Potravinárstvo Slovak Journal of Food Sciences*, 12(1), 566–569. <https://doi.org/10.5219/959>

17. Chernukha, I.M., Kotenkova, E.A. (2018). Influence of functional food product on serum fatty acid composition in hyperlipidemic rats. *Food systems*, 1(4), 4–9. <https://doi.org/10.21323/2618-9771-2018-1-4-4-9> (in Russian)

18. Kotenkova, E.A., Fedulova, L.V., Chernukha, I.M. (2017). The study of isolated from sus scrofa aorta tissue-specific substances with a molecular weight less than 30 kDa. *Vsyo o myase*, 2, 40–42. (in Russian)

19. Fedulova, L.V., Vasilevskaya, E.R., Kotenkova, E.A., Kalinova, E.A. (2018). Algorithm of in vitro assessment for products containing bioactive substances. *Vsyo o myase*, 6, 47–49. <https://doi.org/10.21323/2071-2499-2018-6-47-49> (in Russian)

20. Vasilevskaya, E.R., Kotenkova, E.A., Lukoniva, E.A., Kalinova, E.A. (2017). Research methodology of Sus Scrofa tissue extracts protein-peptide components. *Theory and practice of meat processing*, 2(3), 79–85. <https://doi.org/10.21323/2414-438X-2017-2-3-79-85> (in Russian)

21. Li, L., Liu, Y., Zhou, G., Xu, X., Li, C. (2017). Proteome Profiles of Digested Products of Commercial Meat Sources. *Frontiers in Nutrition*, 4, 8. <https://doi.org/10.3389/fnut.2017.00008>

22. Wen, S., Zhou, G., Song, S., Xu, X., Voglmeir, J., Liu, L., Zhao, F., Li, M., Li, L., Yu, X., Bai, Y., Li, C. (2015). Discrimination of in vitro and in vivo digestion products of meat proteins from pork, beef, chicken, and fish. *Proteomics*, 15(21), 3688–3698. <https://doi.org/10.1002/pmic.201500179>

AUTHOR INFORMATION

Ekaterina R. Vasilevskaya — candidate of technical sciences, researcher, Experimental clinic-laboratory of biologically active substances of animal origin, V. M. Gorbатов Federal Research Center for Food Systems of Russian Academy of Sciences. 109316, Moscow, Talalikhina str., 26. Tel.: +7–495–676–92–11, E-mail: e.vasilevskaya@fnpcs.ru

ORCID: <https://orcid.org/0000-0002-4752-3939>

* corresponding author

Anastasiya G. Akhremko — junior researcher, Experimental clinic-laboratory of biologically active substances of animal origin, V. M. Gorbатов Federal Research Center for Food Systems of Russian Academy of Sciences. 109316, Moscow, Talalikhina str., 26. Tel.: +7–495–676–92–11, E-mail: a.akhremko@fnpcs.ru

ORCID: <https://orcid.org/0000-0002-0211-8171>

Ekaterina K. Polishchuk — senior laboratory assistant, Experimental clinic-laboratory of biologically active substances of animal origin, V. M. Gorbатов Federal Research Center for Food Systems of Russian Academy of Sciences. 109316, Moscow, Talalikhina str., 26. Tel.: +7–495–676–92–11, E-mail: e.polishchuk@fnpcs.ru

ORCID: <https://orcid.org/0000-0003-2719-9649>

Liliya V. Fedulova — candidate of technical sciences, Head of Experimental clinic-laboratory of biologically active substances of animal origin, V. M. Gorbатов Federal Research Center for Food Systems of Russian Academy of Sciences. 109316, Moscow, Talalikhina str., 26. Tel.: +7–495–676–92–11, E-mail: l.fedulova@fnpcs.ru

ORCID: <https://orcid.org/0000-0003-3573-930X>

All authors bear responsibility for the work and presented data.

All authors made an equal contribution to the work.

The authors were equally involved in writing the manuscript and bear the equal responsibility for plagiarism.

The authors declare no conflict of interest.

Received 08.08.2020 Accepted in revised 15.09.2020 Accepted for publication 25. 09.2020

THE CHANGES IN THE AMINO AND FATTY ACID PROFILES IN THE SEMIFINISHED FOODSTUFFS BASED ON BROILER MEAT AND COMPONENTS OF CHICKEN EGGS AFTER DIFFERENT TYPES OF THERMAL TREATMENT

Isabella L. Stefanova*, Liudmila V. Shakhnazarova, Anastasia Yu. Klimenkova, Inna M. Sorokina

All-Russian Scientific Research Institute of Poultry Processing Industry — Branch of the Federal State Budget Scientific Institution Federal Scientific Center “All-Russian Research and Technological Poultry Institute” of Russian Academy of Sciences, Rzhavki township, Moscow region

Keywords: broiler meat, coagulated chicken egg melange, semifinished foodstuff, thermal treatment, amino and fatty acid profiles, nutrient balance

Abstract

The changes in the amino and fatty acid profiles in the semifinished foodstuffs (SFFs) based on broiler meat and coagulated chicken egg melange after different types of thermal treatment (water or steam boiling, braising, baking, frying) were studied. The amino acid profiles were determined on Knauer analyzer; tryptophan by standard method. The biological value of the treated products was assessed using amino acid balance coefficients calculated by the method of N. N. Lipatov. It was found that the changes in the initial amino acid profiles of the SFFs were the least after water and steam boiling; braising and baking were found to increase the contents of the essential amino acids. The amino acid profiles in the treated SFFs were close to the reference values. The best criteria of their biological value (coefficient of rationality of amino acid composition, comparable redundancy) were found after water and steam boiling. It was found that all types of thermal treatments insignificantly affected the parameters of fatty acid balance within the SFFs; the changes found were primarily related to slight increase in total content of saturated fatty acids and increase in total content of polyunsaturated fatty acids (PUFAs) in compare to initial profiles, by 2.64–3.88% depending on the treatment type. The changes in ω -6/ ω -3 PUFAs ratios were more substantial especially after braising.

Introduction

The development and reinforcement of the health status in human of different ages cannot be achieved without consumption of the foodstuffs with high content of essential nutrients (well-balanced protein and fat, etc.). The increasing demand for these foodstuffs is a constant trend of the food market worldwide.

The everyday consumption of poultry meat can provide consumers with vitally important essential amino acids (tryptophan, lysine, and methionine) in the most beneficial ratio 1:3:3 [1,2]; these amino acids take part in the biosynthesis of different tissue proteins and also play certain special functions in human. However, the protein of broiler meat is known to be valine-deficient and hence the foodstuffs fully balanced for all essential amino acids should be additionally corrected for the valine content since the deficit (or absence) of this amino acid can result in the severe dysfunctions of the central nervous system and myasthenia [3].

Another important aspect of healthy nutrition is the balance in the fatty acid profile of the dietary lipids since saturated (SFA), monounsaturated (MUFA), and polyunsaturated (PUFA) fatty acids are the main structural and functional components of cell membranes. The special attention should be paid to the essential PUFAs which cannot be synthesized by animals and human and should be provided via feed or food, primarily linolic (LA, C18:2,

ω -6) and α -linoleic (ALA, C18:3, ω -3) acids. These PUFAs are the precursors of a wide range of different eicosanoids, hormone-like substances protecting and maintaining structural and functional integrity of cells and cell components [4,5,6]. The ω -3 and ω -6 PUFAs are the necessary dietary nutrients for all vertebrates since their presence and ratio influence the status of lipid metabolism, susceptibility to the cardiovascular diseases, disturbances in neural and ophthalmic functions, allergic diseases, inflammations [5,7,8,9,10]. On the gene level these PUFAs control gene expression in different organs [8,9] and tissues [11,12,13]. The substantial part (ca. 50–70%) of dietary PUFAs is catabolyzed to meet energy requirements in human [14,15]. Only minor part of the dietary essential PUFAs undergoes the biotransformation to eicosahexaenic and docosahexaenic acids [4,16], main fatty acids in the retinal membranes and the precursors of local cell-regulating hormones affecting the inflammation, bloodstream regulation, prepartum fetal loss, etc. [17].

Poultry meat contains large amounts of the PUFAs; however, the ω -6 acids are predominant and the ω -6/ ω -3 PUFAs ratio is relatively large (12:1 in lipids of broiler meat and 25:1 in turkey) in compare to the recommended level (no more than 1:10).

With all aforementioned taken into account, the development of new technologies or modification of the existing

FOR CITATION:

Stefanova I. L., Shakhnazarova L. V., Klimenkova A.YU., Sorokina I. M. The changes in the amino and fatty acid profiles in the semifinished foodstuffs based on broiler meat and components of chicken eggs after different types of thermal treatment. *Theory and practice of meat processing*. 2020; 5(3): 22–27. <https://doi.org/10.21323/2414-438X-2020-5-3-22-27>.

foodstuffs involving the additional supplementation with ingredients with high biological and nutritive value is as urgent task for modern food industry.

The enrichment of foodstuffs with protein and fat ingredients usually involves the supplementation with collagen-rich animal meat products [18] and butter or vegetable (mostly sunflower) oils, respectively. All these ingredients are rich in ω -6 PUFAs and the resulting dietary ω -6/ ω -3 PUFAs ratio is far from optimal. The use of the ingredients based on poultry eggs and their components can be the valuable alternative since these ingredients could be the source of high-quality protein containing almost all essential amino acids [19]. The content of valine and some other essential amino acids in chicken eggs is higher in compare to protein of broiler meat.

Egg lipids contain simple fats, phospholipids (including lecithin, cephalin, sphingomyelin), cerebrosides, sterides, sterines (cholesterol). Choline presents up to 75% of egg phospholipids; concentration of lecithin is 6-fold higher in compare to cholesterol promoting the better bioavailability of egg lipids. However, the most value ingredients within the eggs are essential PUFAs deficient in other standard foodstuffs in human diets.

Eggs also have certain important technological properties like foam and gel production, emulsifying ability, ability to make a product "airy". The most important technological factor is the gel producing ability of the albumen [20].

The coagulation of egg components is a prospective technology of maximal preservation of the useful properties of the eggs. The studies on the changes in the latter induced by different factors resulted in the development of protein-rich food ingredients based on the coagulated egg components (melange, albumen, yolk) with grainy texture [21]. The use of these ingredients as the additives to meat can increase the contents of full-value protein and fat in standard and functional food commodities to enhance their biological value.

The use of chopped meat-based semifinished foodstuffs (SFFs) in human diets requires the knowledge on the changes in nutritional and biological value which occur in the SFFs during the last stage of cooking, different types of thermal treatment. These types are known to differently affect the composition of the SFFs [21,22]. However, there is a lack of available data on the changes in the biological value of the SFFs after the thermal treatment; this knowledge is necessary for the formulation of diets for different categories and ages of consumers, receipts of special and functional nutrition. There is also a lack of information regarding the effects of thermal treatments on the combined SFFs containing poultry meat and egg-based ingredients.

The aim of the study presented was the investigation of the effects of different types of thermal treatment (cooking) on the parameters of biological value of the combined SFF containing broiler meat and coagulated chicken egg melange (CCEM).

Materials and methods

The SFF in this study was the cutlet-like semifinished product contained chopped broiler meat (55%), CCEM (20%), milk powder (12%), wheat bread (9%), dried onion (3%), salt (0.8%), and spices.

The CCEM was preliminary produced via the heating of acidified melange of chicken eggs until the formation of a clot with the texture similar to grainy curd [23]. The yield of CCEM was 82.7% of initial melange; CCEM produced contained 14.3% of protein and 12.1% of fat.

The types of the thermal treatment used for the cooking of the SFF included water boiling (water/SFF ratio 0.5:1.0), steam boiling, traditional techniques of braising, baking, and frying; the amino and fatty acid profiles of cooked SFF were compared to the respective parameters before the treatments.

The amino acid profiles were determined on Knauer analyzer; tryptophan by standard method according to Russian standard GOST 34132. The biological value of the protein within the treated products was assessed using amino acid balance coefficients calculated by the method of N. N. Lipatov [24]. The following criteria were used: amino acid score (AS); coefficient of rationality of amino acid composition (Rc); "comparable redundance" of essential fatty acids (σ), utilitarian coefficient (α_j).

Fatty acid profiles were determined via capillary gas-liquid chromatography. The total amounts of SFAs, MUFAs, and PUFAs were determined; their ratios and ω -6/ ω -3 PUFAs ratios were calculated.

The percentages of moisture, protein, and fat in the products were determined according to GOST 31470.

The temperature of the thermal treatments was controlled by the thermometer with the scale from -50 to $+300$ °C and maximum relative measurement error ± 0.5 °C.

Results and discussion

The preliminary comparative investigation of the amino and fatty acid profiles in the intact SFF with CCEM in compare to the SFF without CCEM supplementation has revealed the significant increases in the contents of essential amino acids: methionine by 35%, valine and isoleucine by 22–23%, threonine, leucine, phenylalanine, and lysine by 14–15% while the content of tryptophan in the CCEM-supplemented SFF has been slightly lower; the ratio of essential ω -6/ ω -3 PUFAs has been better in the CCEM-supplemented SFF. The conclusion has been made that the balance of the amino and fatty acids has been better in the supplemented product. The σ criterion has been lower in the CCEM-supplemented SFF while the Rc criterion has been higher; these findings have evidenced the better biological value of this product.

The comparative investigation of the amino and fatty acid profiles in the CCEM-supplemented SFF after different types of thermal treatment (cooking) in compare to the intact one revealed that all treatments affected the contents of moisture, fat, and protein (Table 1).

Table 1. The changes in basic nutritive parameters in the SFF depending on cooking (thermal treatment) type

Treatment type	Content, %		
	Moisture	Fat	Protein
Raw (untreated)	70.9	6.7	14.3
Water boiled	70.9	6.3	14.5
Steam boiled	69.4	6.5	15.3
Braised	69.6	6.4	15.3
Baked	62.7	10.0	18.1
Fried	65.2	10.3	16.1

The moisture content underwent little changes during the thermal treatments. Moisture loss after braising, steam boiling, and frying was 1.3; 1.5 and 5.7%, respectively; the highest moisture loss was found for baking (8.2%).

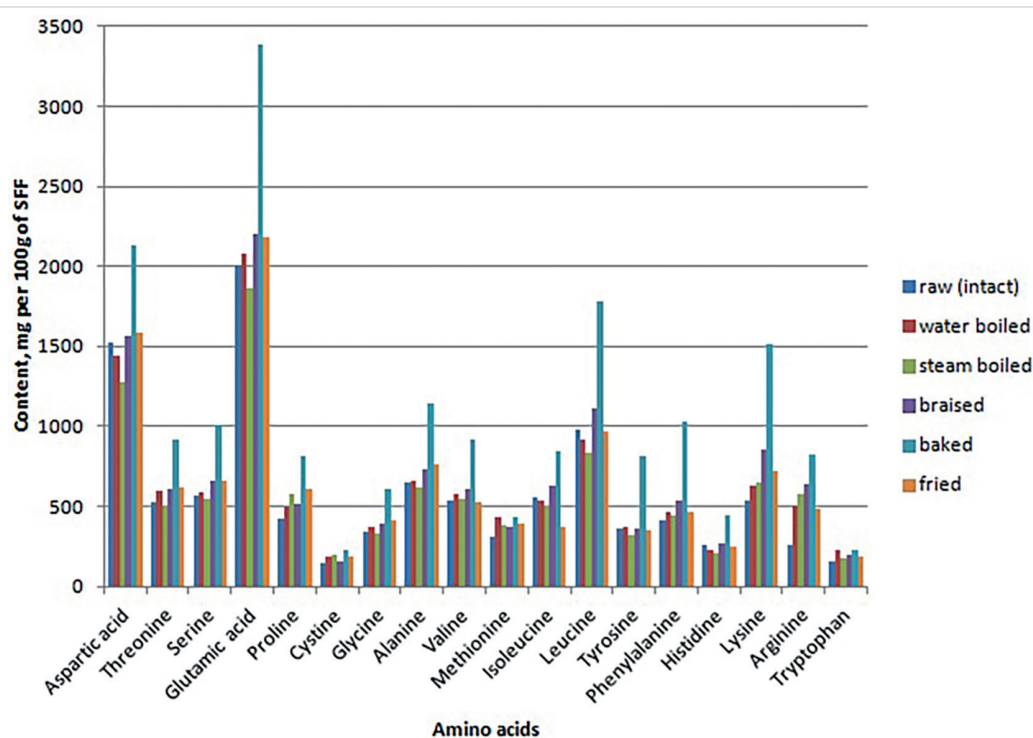
The thermal treatments resulted in the increases in protein content within the cooked SFF: baking by 3.8%, frying by 1.8%, braising and steam boiling by 1.0%, water boiling by 0.2%.

The changes in fat content in cooked SFF were different: braising, water and steam boiling resulted in the slight decreases in fat content while baking and frying increased this parameter by 3.0 and 3.3%, respectively.

The study of amino acid profiles of the proteins of intact and cooked SFF revealed the trend to higher contents of essential amino acids after all types of cooking (Figure 1).

The limiting amino acid in raw SFF was lysine with AS criterion 0.68 units. Different types of thermal treatment resulted in different changes in the amino acid profiles of protein within the SFF. The least affected amino acid profiles (in compare to the raw SFF) were found for water and steam boiling. Braising and baking resulted in higher contents of essential amino acids; AS for essential amino acids in these cases were close to the reference levels (close to 1.0) with the exception of tryptophan (AS1.82). No limiting amino acids were found in baked SFF though AS for certain essential amino acids (leucine, lysine, threonine, tryptophan, phenylalanine, and tyrosine) were substantially higher than 1.0 (Table 2). The utilitarian coefficients α_j (the content of essential amino acids in relation to the respective physiologic requirements of consumer) in the cooked SFF were close to the intact one.

The calculated criteria of biological value (coefficient of rationality of amino acid composition R_c and “comparable

**Figure 1.** Amino acid profiles of the SFF depending on the type of the thermal treatment**Table 2.** The essential amino acid scores (AS) and utilitarian coefficients (α_j) in raw and cooked SFF

Amino acids	Raw		Water boiled		Steam boiled		Braised		Baked		Fried	
	AS	α_j	AS	α_j	AS	α_j	AS	α_j	AS	α_j	AS	α_j
Valine	0.76	0.89	0.79	1.00	0.72	1.00	0.80	1.00	1.02	1.00	0.65	0.89
Isoleucine	0.98	0.69	0.92	0.86	0.83	0.87	1.03	0.78	1.17	0.87	0.58	1.00
Leucine	0.98	0.69	0.91	0.87	0.78	0.92	1.04	0.77	1.41	0.72	0.86	0.67
Lysine	0.68	1.00	0.79	1.00	0.78	0.92	1.02	0.78	1.52	0.67	0.82	0.71
Methionine+cystine	0.91	0.75	1.23	0.64	1.08	0.67	0.98	0.82	1.03	0.99	1.42	0.41
Threonine	0.92	0.74	1.03	0.77	0.83	0.84	1.00	0.80	1.26	0.81	0.96	0.60
Tryptophan	1.11	0.61	1.54	0.51	1.19	0.61	1.82	0.44	1.29	0.79	1.15	0.50
Phenylalanine+tyrosine	0.90	0.76	0.96	0.82	0.83	0.87	0.98	0.82	1.70	0.60	0.85	0.68

redundance" of essential fatty acids σ) showed that water and steam boiling resulted in the best parameters of biological value of protein within the FFS (Table 3).

Table 3. The criteria of biological value of raw and cooked SFF

Treatment type	Coefficient of rationality of amino acid composition Rc	"Comparable redundancy" of essential fatty acids σ	Biological value
Raw (untreated)	0.77	10.61	77.5
Water boiled	0.83	7.12	76.9
Steam boiled	0.86	5.57	84.0
Braised	0.80	9.04	71.6
Baked	0.76	11.17	70.7
Fried	0.67	17.79	66.9

The study of the individual fatty acid profiles in the cooked CCEM-supplemented SFF (Table 4) and total contents of SFAs, MUFAs, and PUFAs (Table 5) revealed that water boiling, steam boiling, braising, and baking slightly increase the total SFA content (by 1.37; 1.89; 4.32 and 2.29%, respectively); the most substantial changes were found after braising. The total contents of MUFAs with these treatment types were similar to the level of the raw product.

Table 4. Fatty acid (FA) profiles in raw and cooked SFF

FA	FA type	Content, % of total FA					
		Raw	Water boiled	Steam boiled	Braised	Baked	Fried
Butyric	C4:0	0.13	0.10	0.09	0.13	0.09	0.12
Capronic	C6:0	0.09	0.07	0.07	0.08	0.07	0.08
Caprylic	C8:0	0.06	0.05	0.05	0.06	0.05	0.06
Caprinic	C10:0	0.14	0.13	0.14	0.15	0.14	0.12
Lauric	C12:0	0.17	0.18	0.18	0.18	0.17	0.14
Tridecanoic	C13:0	0.03	0.03	0.03	0.13	0.03	0.05
Myristinic	C14:0	0.89	0.93	0.97	1.02	0.94	0.75
Myristoleic	C14:1	0.14	0.14	0.13	0.07	0.14	0.10
Pentadecanoic	C15:0	0.17	0.17	0.20	0.47	0.18	0.22
Palmitic	C16:0	22.43	23.53	24.20	25.49	24.12	19.42
Palmitoleic	C16:1	3.93	3.93	3.94	3.77	3.85	3.11
Margaric	C17:0	0.14	0.14	0.15	—	0.15	0.11
Margaroleic	C17:1	0.05	0.05	0.06	—	0.07	0.05
Stearic	C18:0	6.50	6.79	6.96	7.67	7.11	6.07
Oleic	C18:1	35.33	36.31	36.71	36.31	35.52	32.95
Elaidinic	C18:1(t9)	1.62	1.64	1.66	1.57	1.63	1.39
Linolic	C18:2(n6)	25.76	24.24	23.06	22.28	23.32	33.72
γ -linoleic	C18:3(n6)	0.18	0.13	0.13	—	0.12	0.15
α -linoleic	C18:3(n3)	0.42	0.35	0.32	0.28	0.33	0.33
Arachidonic	C20:0	0.08	0.08	0.09	—	0.09	0.11
Gondoic	C20:1	0.34	0.38	0.38	—	0.38	0.35
Eicosatrienoic	C20:3	0.43	0.41	0.31	0.34	0.33	0.30
Geneicosanoic	C21:0	0.22	0.23	0.20	—	0.20	0.14
Begenic	C22:0	—	—	—	—	—	0.18
Docosenoic	C22:1	0.72	—	—	—	—	—

Table 5. The contents of total SFAs, MUFAs, and PUFAs in raw and cooked SFF, % of total FA

Total	Cooking type					
	Raw	Water boiled	Steam boiled	Braised	Baked	Fried
SFAs	31.05	32.42	33.34	35.37	33.34	27.56
MUFAs	42.16	42.45	42.83	41.72	41.59	37.94
PUFAs	26.79	25.13	23.83	22.90	24.10	34.49

The slight changes in the fatty acid profiles in cooked SFF were primarily related to the decreases in the PUFAs contents; in water-boiled, steam-boiled, braised and baked SFF this decrease was 2.64; 2.96; 3.88 and 2.68%, respectively. The ω -6/ ω -3 PUFAs ratios were affected more substantially especially in the case of braising; the least change in this ratio was found in water-boiled SFF. Generally, all types of cooking (thermal treatments) had little influence on the fatty acid balance.

Conclusion

The amino and fatty acid profiles in the SFF supplemented with coagulated chicken egg melange after different types of cooking (thermal treatments) were studied.

Different types of cooking differently affected the contents of protein, fat, amino and fatty acids within the product.

The least changes in the parameters of biological and nutritive value of the SFF was found with water- and steam-boiling. Braising and baking were found to increase the contents of essential amino acids; the resulting contents of the latter were close to the respective reference levels (AS close to 1.0) with the exception of tryptophan (AS1.82). No limiting amino acids were found in baked SFF though AS for certain essential amino acids (leucine, lysine, threonine, tryptophan, phenylalanine, and tyrosine) were substantially higher than 1.0 (i. e. higher in compare to the reference levels).

The highest biological value (as revealed by the coefficient of rationality of amino acid composition Rc and "comparable redundancy" of essential fatty acids σ) had steam-boiled SFF; overall index of biological value for this treatment was the highest (84.0) in compare to the raw SFF (77.5).

The effects of different types of thermal treatment on the fatty acid profiles of SFF were insignificant. The slight changes in the fatty acid profiles in cooked SFF were primarily related to the increases in the SFAs and decreases in the PUFAs contents; in water-boiled, steam-boiled, braised and baked SFF this decrease was 2.64; 2.96; 3.88 and 2.68%, respectively. The ω -6/ ω -3 PUFAs ratios were affected more substantially especially in the case of braising.

REFERENCES

1. Tutelyan, V.A. (2000). On the correction of micronutrient deficits for the improvement of nutrition and health of infant and adult population on the threshold of the third millennium. *Voprosy pitaniya*, 4, 6–7. (in Russian)
2. Pokrovsky, A.A. (1994). Biochemical substantiation of the development of the foodstuffs with increased biological value. *Voprosy Pitaniia*, 1, 1–3. (in Russian)
3. Erastov, G.M. (2014). The nutritional value of poultry meat. *Pitisevodstvo*, 3, 28–30. (in Russian)
4. Wall, R., Ross, R.P., Fitzgerald, G.F., Stanton, C. (2010). Fatty acids from fish: the anti-inflammatory potential of long-chain omega-3 fatty acids. *Nutrition Reviews*, 68(5), 280–289. <https://doi.org/10.1111/j.1753-4887.2010.00287.x>
5. Vasilyev, A.V., Sharanova, N.E., Kulakova, S.N. (2014). Nutrimetabolomics — the new stage of biochemistry of nutrition. The role of nutr lipidomic analysis. *Voprosy Pitaniia*, 83(1), 4–11. (in Russian)
6. Farooqui, A.A. (2012). n-3 fatty acid-derived lipid mediators in the brain: new weapons against oxidative stress and inflammation. *Current Medicinal Chemistry*, 19(4), 532–543. <https://doi.org/10.1016/j.pain.2007.01.020>
7. Golberg, R.J., Katz, J. (2007). A meta-analysis of the analgesic effects of omega-3 polyunsaturated fatty acid supplementation for inflammatory joint pain. *Pain*, 129(1), 210–223. <https://doi.org/10.1016/j.pain.2007.01.020>
8. Leaf, A. (2006). Prevention of sudden cardiac death by n-3 polyunsaturated fatty acids. *Fundamental and Clinical Pharmacology*, 20(6), 525–538. <https://doi.org/10.1111/j.1472-8206.2006.00438.x>
9. SanGiovanni, J.P., Chew, E.Y. (2005). The role of omega-3 long-chain polyunsaturated fatty acids in health and disease of the retina. *Progress in Retinal and Eye Research*, 24(1), 87–138. <https://doi.org/10.1016/j.preteyeres.2004.06.002>
10. Uauy, R., Dangour, A.D. (2006). Nutrition in brain development and aging: role of essential fatty acids. *Nutrition Reviews*, 64, S24–S33. <https://doi.org/10.1111/j.1753-4887.2006.tb00242.x>
11. Barceli-Coblijn, G., Hogg, E., Kitajka, K., Puskas, L.G., A. Zvara, A., Hackler, L., Nyakas, C., Penke, Z., Farkas, T. (2003). Modification by docosahexaenoic acid of age-induced alterations in gene expression and molecular composition of rat brain phospholipids. *Proceedings of the National Academy of Sciences*, 100(20), 11321–11326. <https://doi.org/10.1073/pnas.1734008100>
12. Puskas L. G., Kitajka K., Nyakas, Barcelo-Coblijn, G., Farkas, T. (2003). Short-term administration of omega-3 fatty acids from fish oil results in increased transthyretin transcription in old rat hippocampus. *Proceedings of the National Academy of Sciences*, 100(4) 6 1580–1585. <https://doi.org/10.1073/pnas.0337683100>
13. Clarke, S.D. (2001). Polyunsaturated fatty acid regulation of gene transcription: a molecular mechanism to improve the metabolic syndrome. *The Journal of Nutrition*, 131(4), 1129–1132. <https://doi.org/10.1093/jn/131.4.1129>
14. Duplus, E., Glorian, M. (2000). Fatty acid regulation of gene transcription. *Journal of Biological Chemistry*, 275(40), 30749–30752. <https://doi.org/10.1074/jbc.R000015200>
15. Wahle, K.W.J., Rotondo, D., Heys, S.D. (2003). Polyunsaturated fatty acids and gene expression in mammalian systems. *Proceedings of the Nutrition Society*, 62(2), 349–360. <https://doi.org/10.1079/pns2003249>
16. Broadhurst, C.L., Wang, Y., Crawford, M.A., Cunnane, S.C., Parkington, J.E., Schmidt, W.F. (2002). Brain-specific lipids from marine, lacustrine, or terrestrial food resources: potential impact on early African Homo sapiens. *Comparative Biochemistry and Physiology Part B: Biochemistry and Molecular Biology*, 131(4), 653–673. [https://doi.org/10.1016/s1096-4959\(02\)00002-7](https://doi.org/10.1016/s1096-4959(02)00002-7)
17. Umegaki, K., Hashimoto, M., Yamasaki, H., Fujii, Y., Yoshimura, M., Sugisawa, A., Shinozuka, K. (2001). Docosahexaenoic acid supplementation-increased oxidative damage in bone marrow DNA in aged rats and its relation to antioxidant vitamins. *Free Radical Research*, 34(4), 427–435. <https://doi.org/10.1080/10715760100300361>
18. Davis, B.C., Kris-Etherton, P.M. (2003). Achieving optimal essential fatty acid status in vegetarians; current knowledge and practical implications. *The American Journal of Clinical Nutrition*, 78(3), 640S–646S. <https://doi.org/10.1093/ajcn/78.3.640s>
19. Omarov, R.S., Sycheva, O.V., Shlykov, S.N. (2011). Animal proteins in the technologies of meat products. *Meat branch*, 3(99), 36–38. (in Russian)
20. Stefanova, I.L., Mazo, V.K., Mokshantseva, I.V., Klimenkova, A. Yu. (2017). The perspectives of the use of egg albumen within functional food ingredients. *Poultry and Chicken Products*, 1, 43–45. (in Russian)
21. Stefanova, I.L., Shakhnazarova, L.V., Klimenkova, A. Yu. (2019). Technology development for eggs and egg components further processing and its introduction to industry. *Proceedings scientific and technical support efficiency and quality production of agricultural products*. 246–257. <https://doi.org/10.30975/978-5-9909889-2-7-2019-1-1-246-257> (in Russian)
22. Nys, Y., Bain, M., Immerseel, F.V. (2011). Improving the Safety and Quality of Eggs and Egg Products: Vol. 1: Egg Chemistry, Production and Consumption. Woodhead Publishing Limited. — 602 p. ISBN: 9781845697549
23. Stefanova, I.L., Shakhnazarova, L.V., Klimenkova, A. Yu., Krasnyukov, Yu.N. (2017). The investigation of the process of coagulation of egg melange and qualitative characteristics of the resulting product. *Poultry and Chicken Products*, 5, 49–53. (in Russian)
24. Lipatov, N.N., Lisitsyn, A.B., Yudina, S.B. (1996). The advancement of the methods of design of the biological value of foodstuffs. *Storage and processing of farm products*, 2, 24–25. (in Russian)

AUTHOR INFORMATION

Isabella L. Stefanova — doctor of technical sciences, chief researcher, Laboratory Of Infant And Special Poultry Products, All-Russian Scientific Research Institute of Poultry Processing Industry — Branch of the Federal State Budget Scientific Institution Federal Scientific Center “All-Russian Research and Technological Poultry Institute” of Russian Academy of Sciences, 142552, Moscow region, Rzhavki township. Tel.: +7-495-944-53-30, E-mail: dp.vniipp@mail.ru

ORCID: <https://orcid.org/0000-0002-4394-5149>

*corresponding author

Liudmila V. Shakhnazarova — candidate of technical sciences, leading researcher, Laboratory Of Infant And Special Poultry Products, All-Russian Scientific Research Institute of Poultry Processing Industry — Branch of the Federal State Budget Scientific Institution Federal Scientific Center “All-Russian Research and Technological Poultry Institute” of Russian Academy of Sciences, 142552, Moscow region, Rzhavki township. Tel.: +7-495-944-53-30, E-mail: dp.vniipp@mail.ru

ORCID: <https://orcid.org/0000-0001-5671-6495>

Anastasia Yu. Klimenkova — researcher, Laboratory Of Infant And Special Poultry Products, All-Russian Scientific Research Institute of Poultry Processing Industry — Branch of the Federal State Budget Scientific Institution Federal Scientific Center “All-Russian Research and Technological Poultry Institute” of Russian Academy of Sciences, 142552, Moscow region, Rzhavki township. Tel.: +7-495-944-53-30, E-mail: dp.vniipp@mail.ru

ORCID: <https://orcid.org/0000-0002-3272-9467>

Inna M. Sorokina — researcher, Testing laboratory center, All-Russian Scientific Research Institute of Poultry Processing Industry — Branch of the Federal State Budget Scientific Institution Federal Scientific Center “All-Russian Research and Technological Poultry Institute” of Russian Academy of Sciences, 142552, Moscow region, Rzhavki township. Tel.: +7-495-944-53-30, E-mail: dp.vniipp@mail.ru

ORCID: <https://orcid.org/0000-0001-7859-4011>

All authors bear responsibility for the work and presented data.

All authors made an equal contribution to the work.

The authors were equally involved in writing the manuscript and bear the equal responsibility for plagiarism.

The authors declare no conflict of interest.

Received 25.06.2020 Accepted in revised 15.09.2020 Accepted for publication 28.09.2020

DEVELOPMENT OF HIGH SENSITIVE REAL-TIME PCR TO DETECT MUSTARD AND OTHER ALLERGENS OF THE FAMILY *BRASSICACEAE* IN FOOD SAMPLES

Konstantin A. Kurbakov,* Valentina N. Zhulinkova, Mihail Yu. Minaev

V. M. Gorbato Federal Research Center for Food Systems of Russian Academy of Sciences, Moscow, Russia

Keywords: real-time PCR, *Brassica juncea*, *Brassica nigra*, *Sinapis alba*, *Brassicaceae*, allergens, food products**Abstract**

Mustard is a commonly used condiment including in production of other food products. As mustard is an allergen, it is necessary to control its presence. The development of PCR test-systems for its detection is complicated by the fact that this condiment can be made from seeds of various plant species (*Brassica juncea*, *Brassica nigra*, *Sinapis alba*) of the family *Brassicaceae* that are not closely related. This family includes other plant species such as white cabbage (*Brassica oleracea*) and rapeseed (*Brassica napus*), which can cause the allergic reaction, although seldom. In this connection, many authors use primers specific to many species of this family, including to allergens, to detect mustard. In this work, we used the similar strategy. To increase sensitivity, primers for the mitochondrial COX gene were selected. To increase PCR stability in analysis of deeply processed products, primers were selected for a region with a length of 61 base pair. In the work, the specificity and sensitivity of the developed PCR method was confirmed. Analyses of different products, including those that underwent deep technological processing, were carried out with these primers. Also, primers were selected to detect white mustard (*S. alba*). When analyzing products on the presence of white mustard, characteristic regional preferences were demonstrated: this species is used in manufacturing products mainly in the UK and USA.

Introduction

Mustard is one of the most commonly used condiments. It is used in many food types, such as spice mixtures, condiments, sauces, marinades, including for meat baking, finished meat products and gastronomy products. Mustard can be prepared from plant seeds of different species: brown mustard (*Brassica juncea*), black mustard (*Brassica nigra*) and white mustard (*Sinapis alba*). Irrespective of a species, which seeds were used for production, mustard is an allergen. Information about its content should be indicated on a label according to TR CU022/2011 [1] and Regulation (EU) No 1169/2011 of the European Parliament and of the Council [2]. More than half of patients with an allergy to mustard also have an increased sensitivity to several other plant-derived foods and pollen [3]. It is necessary to note that the family *Brassicaceae* also includes such allergens as rapeseed, cabbage and broccoli.

The main mustard allergens are 2S albumins of white and brown mustard seeds (Sin a 1 and Bra j 1, respectively) [4,5]. These two albumins have the similar structure and immunological properties. Therefore, when studying mustard as an allergen in a product composition, the species origin of raw materials is of no importance. For *S. alba*, several other allergens were revealed: 11S globulin of seeds (Sin a 2) [6], non-specific lipid transfer protein (Sin a 3) and profilin (Sin a 4) [7]. It is not unlikely that new allergens will be established for *B. juncea*. An allergy to pollen of white cabbage (*Brassica oleracea*) is rare [8]. With that, about half of patients showed cross-sensitivity to mustard. In addition, the search for potential allergens of *B. nigra* has been performed [9].

At present, to control the mustard presence in foods, several methods based on enzyme-linked immunosorbent assay (ELISA) [10,11] and polymerase chain reaction (PCR) are used. With that, ELISA methods showed false positive reactions when analyzing rapeseed [10] or egg protein and soy [11]. When developing PCR methods, the authors propose primer pairs for detection of individual species *B. juncea* and *S. alba*, as well as multiplex test-systems to detect these species together with other allergens. For example, Palle-Reisch *et al.* selected a primer pair for detection of *B. juncea* and *B. nigra* [12]. However, the reaction efficiency was different (100.6% and 85.3%, respectively) when studying DNA of each of these species. In this connection, the limit of detection for *B. nigra* was lower (0.005% DNA in a sample). In another work, Palle-Reisch *et al.* proposed a primer-probe system for the duplex real-time PCR assay for detection of all three mustard species [13]. Sensitivity of this system was 0.0005%. The use of multiplex test-systems is in high-demand for detection of allergens. For example, Köppel *et al.* proposed hexaplex real-time PCR to detect allergens including mustard [14]. In this work, primers proposed by Mustorp *et al.* were used, which were specific not only to possible mustard species, but also to rapeseed (*Brassica napus*), cabbage (*B. oleracea*), radish (*Raphanus sativus*). The detection limit of this reaction was 0.0032%.

The models of sausage meat heated at 75–78 °C for 15 min. were used in [12,13]. Nevertheless, mustard is used in the composition of deeply processed products including canned foods and marinades. This impact leads to a significant DNA degradation. One of the approaches that increase stability

FOR CITATION:

Kurbakov K. A., Zhulinkova V. N., Minaev M. Yu. Development of high sensitive real-time PCR to detect mustard and other allergens of the family *Brassicaceae* in food samples. *Theory and practice of meat processing*. 2020; 5(3): 28–31. <https://doi.org/10.21323/2414-438X-2020-5-3-28-31>

of a PCR method in analysis of degraded DNA is reducing a length of the obtained amplicon [15].

Taking into account the presence of several allergens in the family *Brassicaceae*, impossibility to select a DNA region specific only to species *B. juncea*, *B. nigra* and *S. alba*, and the experience of several researchers, we proposed a primer pair specific to several species of this family. To increase PCR reliability in analysis of non-degraded DNA, primers were selected for a region with a length of 61 base pair.

It is considered that mainly species of the genus *Brassica* are used in Europe and Asia for mustard production, while *Sinapis alba* is used in the USA and Canada. In this work, we also assessed the share of mustard produced using seeds of *Sinapis alba* on the Russian market.

Materials and methods

Objects of investigation

Seeds of white mustard, brown mustard and white cabbage were taken as positive controls. Soy, corn and wheat were used as the negative control.

Samples of mustard available on the Russian market at retail and produced in Russia (3), the Netherlands (1), Germany (1), Poland (1), France (1), USA (3) and UK (1) were studied. The following samples of products containing mustard seeds were taken: marinated vegetables (3), sauce (3), baking chicken marinade (1) and mustard oil (1). Mustard seeds and brine were taken from samples of marinated vegetables.

Sampling and DNA extraction

Seeds were ground in a homogenizer LB20E (Waring Commercial, Torrington, Connecticut, USA). When necessary, food products were minced in a homogenizer GRINDOMIX GM 200 (Retsch, Haan, Germany). Mustard oil (5 ml) was settled for one hour in a centrifuge SIGMA 3-18K (Sigma Zentrifugen, Osterode am Harz, Germany) with acceleration of 29700g at a temperature of 10 °C. After that, the supernatant was removed and sediment was mixed in the 20% Tween 80 solution. For DNA extraction, 100 mg of food products and control samples were taken. Liquid samples were taken with a volume of 200 µl. DNA extraction was carried out using Sorb-GMO-B kit (Syntol, Moscow, Russia) according to the instruction.

Primer design

The primer pairs Brass_fam and Sin_alb were complementary to the mitochondrial COX gene and retrotrans-

poson Sal-T1 (Table 1), respectively. The choice of the mitochondrial DNA region for identification of the family *Brassicaceae* was conditioned by its multi-copy nature, which enhanced sensitivity of the method. The DNA regions were available in the GenBank database [16]. For system design, the programs Primer-BLAST [17] and Oligo-Analyzer v. 3. were used [18].

Conditions for real-time PCR

Real-time PCR was carried out using an amplifier ANK-32 (Syntol, Moscow, Russia). The reaction mixture (30 µl) contained primers with a concentration of 300 nM, 2.5 mM MgCl₂, dNTPs with a concentration of 0.25 mM each, SynTaq polymerase with a concentration of 2.5 activity units and 5 µL of extracted DNA. The components of the reaction mixture were produced by Syntol. The parameter of PCR was as follows: initial denaturation at 95 °C for 7 min and 45 cycles of amplification (60 °C, 40 s and 95 °C, 15 s). All samples were investigated in triplicate. The obtained data were analyzed using the software ANK-32 (Syntol).

Based on the results of PCR with primers Brass_fam, semi-quantitative analysis of the mustard content in the test samples was carried out by the equation:

$$x = E^{(Cq_k - Cq_r)} \quad (1),$$

where

E is the reaction efficiency;

Cq_k is the threshold cycle of the amplification curve of the positive control;

Cq_r is the threshold cycle of the amplification curve of the sample.

Data analysis was carried out using Microsoft Excel 2016 [19].

Results and discussion

Detection of efficiency, specificity and cut-off cycles of positive results

The short length of the PCR amplicon was used for increasing method stability when analyzing samples with degraded DNA.

To detect the reaction parameters and limit of detection, PCR with the selected primer pairs was carried out.

For analysis of primers for the Sin_alb gene, DNA of white mustard seeds and its decimal dilutions of up to 0.0001% were used. For analysis of primers Brass_fam, we used a dilution of DNA of white mustard seeds with a concentration of 100% to 0.0001%. The detection limit of PCR

Table 1. Sequences and positions of the primers used in the study

Primer	Primer sequence (5'-3')	Amplification region	Amplification size, bp
Brass_fam-F	GCCGAGATCAAGGTTCAAACAAA	COX	61
Brass_fam-R	CTTAAATGTCCTTCTTCCCCGC		
Sin_alb-F	GTACGTCTCTAATCGGCATGGAT	Retrotransposn Sal-T1	107
Sin_alb-R	CTGCTGTTCTCTGTTTCGTGAAG		

bp = base pairs

with primers for Brass_fam was 0.001% of the target template in a sample. This value was not lower than in similar test-systems [12,13,14]. The calculated coefficient of correlation of PCR was $R^2 = 0.99$; PCR efficiency was $E = 2.01$. The equation of linear regression is the following:

$$y = -3,317x + 44,97 \quad (2)$$

The calculated coefficient of correlation of PCR with primers Sin_alb was $R^2 = 0.99$; PCR efficiency was $E = 2.03$. The equation of linear regression is the following:

$$y = -3,2x + 51,145 \quad (3)$$

When using Brass_fam and Sin_alb, no non-specific annealing with samples of soy, corn, wheat was found. Non-specific annealing was absent in the reaction between primers Sin_alb and DNA of white cabbage.

Analysis of mustard condiments

The PCR result for the sample of the US organic mustard powder was expectedly close to the Cq values of the control. Relative quantitative analysis of condiments showed that in the samples of domestic mustard, its content was in a range of 0.5 to 4.5% in reference to ground seeds. This value was 0.22%, 1.8% and 13.9% in mustard produced by Germany, the Netherlands and Poland, respectively. The highest content of the mustard powder was in the condiment produced by France (29.6%) and the UK (71%). In one US sample, a mustard proportion was 5.9%, in another 0.004%. Such a low value can be explained by replacement of the mustard powder with some other aromatic raw materials. It is necessary to note that product pH influences the ultimate value of the PCR result [20]. As vinegar is used, as a rule, in production of this condiment, the obtained values can be insignificantly lower than real.

Analysis of other food products

The authors of previous studies tested their methods on different products: cooked sausage models, sauces, spice mixes, meat spreads, fried noodles, nuggets and so on [13,14]. However, there were no deeply processed products among these objects. In our work, we took samples of brine from canned marinated vegetables and mustard oil besides sauces and marinades.

The PCR method allowed detecting mustard in the sauce compositions: 0.6–0.4% of the content relative to the positive control. The mustard content was 10% in the sample of marinade for baking.

The result of the quantitative analysis of the mustard oil composition was 0.005% relative to the positive control,

which is explained by the technology of vegetable oil purification. This result shows that it is impossible to detect mustard oil in the composition of food products using this method. Nevertheless, this method can be used for establishing vegetable oil falsification.

Mustard DNA was revealed in the samples of marinade of canned vegetables at a level of 0.001–0.0015%. With that, it is necessary to note that whole mustard seeds were added. Therefore, it was shown that sensitivity of the developed method allows identifying mustard in the composition of deeply processed products, in particular, canned foods.

Analysis of the use of S. alba seeds in production of ready condiments and other food products

Samples with the mustard content of not lower than 0.1% were used for analysis. It was linked with lower sensitivity of PCR with primers Sin_alb. Whole mustard seeds were taken from the brine of canned products. *S. alba* was qualitatively detected in the products from Poland and the Netherlands as well as USA and UK. In the US and UK mustard samples, mainly white mustard was detected. With that, it should be noted that the condiment sample from the USA was denoted as Braun Mustard. In the European products *S. alba* was detected in lower quantities. It is connected with the fact that these samples belonged to the recipe of “French” mustard, which composition envisages the use of both white mustard and brown or black mustard.

Conclusion

In this work, we proposed a primer pairs to detect mustard in food samples by the PCR method. High sensitivity of the method was shown. This, in particular, allowed detecting mustard traces in deeply processed samples such as canned foods. Also, successful PCR with the sample of DNA extracted from mustard oil was carried out. Therefore, this method can be used for detection of vegetable oil falsification.

Ready condiments and products were compared regarding the use of *S. alba* seeds. Despite availability of raw materials, European manufacturers traditionally use brown and black mustard, while in the UK and USA, white mustard has been detected in condiments declared as brown mustard. Nevertheless, there are condiment types, in which seeds of the genera *Brassica* and *Sinapis* are used. In this connection, it is impossible to rely on the geographical origin of a product and use one primer pair specific only to the genus *Brassica* or *Sinapis*, when analyzing food on the mustard presence.

REFERENCES

1. Technical regulations of the Customs Union TR CU022/2011 “Food products in terms of its labeling” (approved by the decision of the customs Union Commission of December 9, 2011 № 880). Moscow, — 2011. (in Russian)
2. Regulation (EU) No 1169/2011 of 25 October 2011 on the provision of food information to consumers, amending Regulations (EC) No. 1924/2006 and (EC) No. 1925/2006 of the European Parliament and of the Council, and repealing Commission Directive 87/250/EEC, Council Directive 90/496/EEC, Commission Directive 1999/10/EC, Directive 2000/13/EC of the European Parliament and of the Council, Commission Directives 2002/67/EC and 2008/5/EC and Commission Regulation (EC) No 608/2004. *Official Journal of the European Union*, L304, 18–63.

3. Figueroa, J., Blanco, C., Dumpiérrez, A.G., Almeida, L., Ortega, N., Castillo, R., Navarro, L., Pérez, E., Gallego, M.D., Carrillo, T. (2005). Mustard allergy confirmed by double-blind placebo-controlled food challenges: clinical features and cross-reactivity with mugwort pollen and plant-derived foods. *Allergy*, 60(1), 48–55. <https://doi.org/10.1111/j.1398-9995.2005.00644.x>
4. Menéndez-Arias, L., Moneo, I., Domínguez, J., Rodríguez, R. (1988). Primary structure of the major allergen of yellow mustard (*Sinapis alba* L.) seed, Sin a I. *European Journal of Biochemistry*, 177(1), 159–166. <https://doi.org/10.1111/j.1432-1033.1988.tb14357.x-i2>
5. de la Peña, M. A. G., Menéndez-Arias, L., Monsalve, R. I., Rodríguez, R. (1991). Isolation and Characterization of a Major Allergen from Oriental Mustard Seeds, Bra j1. *International Archives of Allergy and Immunology*, 96(3), 263–270. <https://doi.org/10.1159/000235505>
6. Palomares, O., Cuesta-Herranz, J., Vereda, A., Sirvent, S., Villalba, M., Rodríguez, R. (2005). Isolation and identification of an 11S globulin as a new major allergen in mustard seeds. *Annals of Allergy, Asthma & Immunology*, 94(5), 586–592. [https://doi.org/10.1016/S1081-1206\(10\)61138-6](https://doi.org/10.1016/S1081-1206(10)61138-6)
7. Angelina, A., Sirvent, S., Palladino, C., Vereda, A., Cuesta-Herranz, J., Eiwegger, T., Rodríguez, R., Breiteneder, H., Villalba, M., Palomares, O. (2016). The lipid interaction capacity of Sin a 2 and Ara h 1, major mustard and peanut allergens of the cupin superfamily, endorses allergenicity. *Allergy*, 71(9), 1284–1294. <https://doi.org/10.1111/all.12887>
8. Hermanides, H. K., Laheij-de Boer, A. M., Zuidmeer, L., Guikers, C., Ree, R., Knulst, A. C. (2006). Brassica oleracea pollen, a new source of occupational allergens. *Allergy*, 61(4), 498–502. <https://doi.org/10.1111/j.1398-9995.2006.01055.x>
9. Khaliq, B., Falke, S., Negm, A., Buck, F., Munawar, A., Saqib, M., Mahmood, S., Ahmad, M.S., Betzel, C., Akrem, A. (2017). SAXS and other spectroscopic analysis of 12S cruciferin isolated from the seeds of *Brassica nigra*. *Journal of Molecular Structure*, 1137, 60–66. <https://doi.org/10.1016/j.molstruc.2017.02.043>
10. Lee, P. W., Hefle, S. L., Taylor, S. L. Lee, P.-W., Hefle, S.L., Taylor, S.L. (2008). Sandwich enzyme-linked immunosorbent assay (ELISA) for detection of mustard in foods. *Journal of food science*, 73(4), T62-T68. <https://doi.org/10.1111/j.1750-3841.2008.00725.x>
11. Koppelman, S. J., Vlooswijk, R., Bottger, G., Van Duijn, G., Van der Schaft, P., Dekker, J., Van Bergen, H. (2007). Development of an enzyme-linked immunosorbent assay method to detect mustard protein in mustard seed oil. *Journal of food protection*, 70(1), 179–183. <https://doi.org/10.4315/0362-028x-70.1.179>
12. Palle-Reisch, M., Wolny, M., Cichna-Markl, M., Hohegger, R. (2013). Development and validation of a real-time PCR method for the simultaneous detection of black mustard (*Brassica nigra*) and brown mustard (*Brassica juncea*) in food. *Food chemistry*, 138(1), 348–355. <https://doi.org/10.1016/j.foodchem.2012.10.055>
13. Palle-Reisch, M., Cichna-Markl, M., Hohegger, R. (2014). Development and validation of a duplex real-time PCR assay for the simultaneous detection of three mustard species (*Sinapis alba*, *Brassica nigra* and *Brassica juncea*) in food. *Food chemistry*, 153, 66–73. <https://doi.org/10.1016/j.foodchem.2013.12.035>
14. Köppel, R., van Velsen-Zimmerli, F., Bucher, T. (2012). Two quantitative hexaplex real-time PCR systems for the detection and quantification of DNA from twelve allergens in food. *European Food Research and Technology*, 235(5), 843–852. <https://doi.org/10.1007/s00217-012-1806-8>
15. Mano, J., Nishitsuji, Y., Kikuchi, Y., Fukudome, S., Hayashida, T., Kawakami, H., Kurimoto, Y., Noguchi, A., Kondo, K., Teshima, R., Takabatake, R., Kitta, K. (2017). Quantification of DNA fragmentation in processed foods using real-time PCR. *Food chemistry*, 226, 149–155. <https://doi.org/10.1016/j.foodchem.2017.01.064>
16. GenBank®. Bethesda, MD, USA: National Center for Biotechnology Information (NCBI), US National Library of Medicine; 2017. [Electronic resource: <http://www.ncbi.nlm.nih.gov/> Access date 14.07.2020]
17. Primer-BLAST. Bethesda, MD, USA: National Center for Biotechnology Information, U. S. National Library of Medicine; 2017. [Electronic resource: <https://www.ncbi.nlm.nih.gov/tools/primer-blast/index.cgi> Access date 14.07.2020]
18. OligoAnalyzer 3.1, Integrated DNA Technologies, Inc., Coralville, IA, USA; 2017. [Electronic resource: <http://eu.idtdna.com/calc/analyzer> Access date 14.07.2020]
19. Microsoft Excel 2010, Microsoft, Redmond, WA, USA; 2016.
20. Bauer, T., Hammes, W. P., Haase, N. U., Hertel, C. (2004). Effect of food components and processing parameters on DNA degradation in food. *Environmental Biosafety Research*, 3(4), 215–223. <https://doi.org/10.1051/eb:2005005>

AUTHOR INFORMATION

Konstantin A. Kurbakov — Engineer of Laboratory of hygiene of manufacture and microbiology, V. M. Gorbатов Federal Research Center for Food Systems of Russian Academy of Sciences. 109316, Moscow, Talalikhina str., 26.

Tel: +7-495-676-63-21. E-mail: homo_ludens@vniimp.ru

<https://orcid.org/0000-0003-1348-860X>

* corresponding author

Valentina N. Zhulinkova — Engineer of Laboratory of hygiene of manufacture and microbiology, V. M. Gorbатов Federal Research Center for Food Systems of Russian Academy of Sciences. 109316, Moscow, Talalikhina str., 26.

Tel: +7-495-676-63-21. E-mail: vzhulinkova@fncpc.ru

<https://orcid.org/0000-0001-9063-2164>

Mihail Yu. Minaev — candidate of technical sciences, head of Laboratory of molecular biology and bioinformatics, V. M. Gorbатов Federal Research Center for Food Systems of Russian Academy of Sciences. 109316, Moscow, Talalikhina str., 26. Tel.: +7-495-676-79-81. E-mail: mmi-naev@inbox.ru

<https://orcid.org/0000-0002-0038-9744>

All authors bear responsibility for the work and presented data.

All authors made an equal contribution to the work.

The authors were equally involved in writing the manuscript and bear the equal responsibility for plagiarism.

The authors declare no conflict of interest.

Received 13.08.2020 Accepted in revised 01.09.2020 Accepted for publication 25.09.2020